UCLA

Statistical Methods for Bulk and Single-cell RNA Sequencing Data

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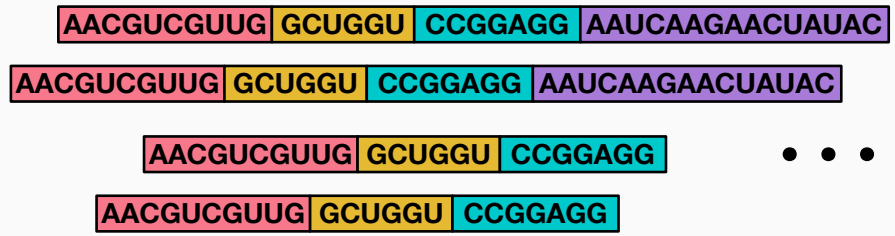
RNA sequencing (RNA-seq) technology

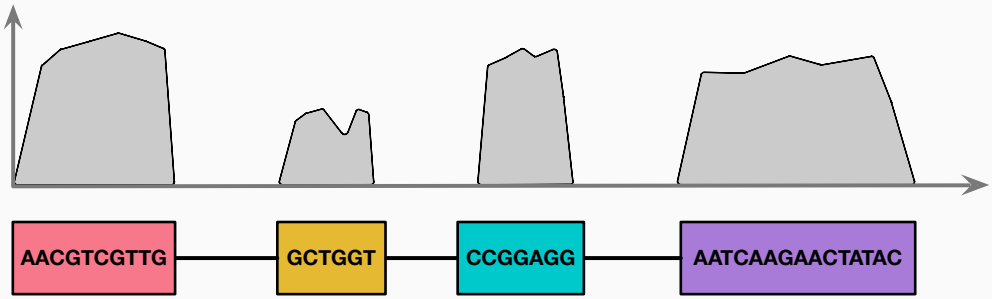
full length RNA isoforms
(unknown)



RNA-seq
experiments

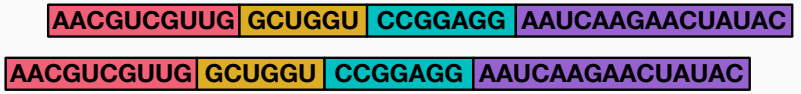
statistical
inference

RNA-seq data
(observed)



RNA sequencing (RNA-seq) experiment

full length RNA isoforms
(1712 bp on average)



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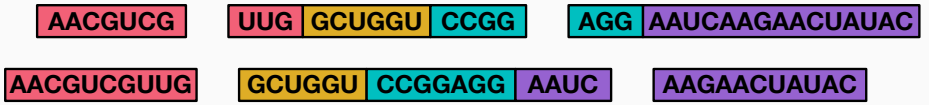
RNA fragments
(< 600 bp)



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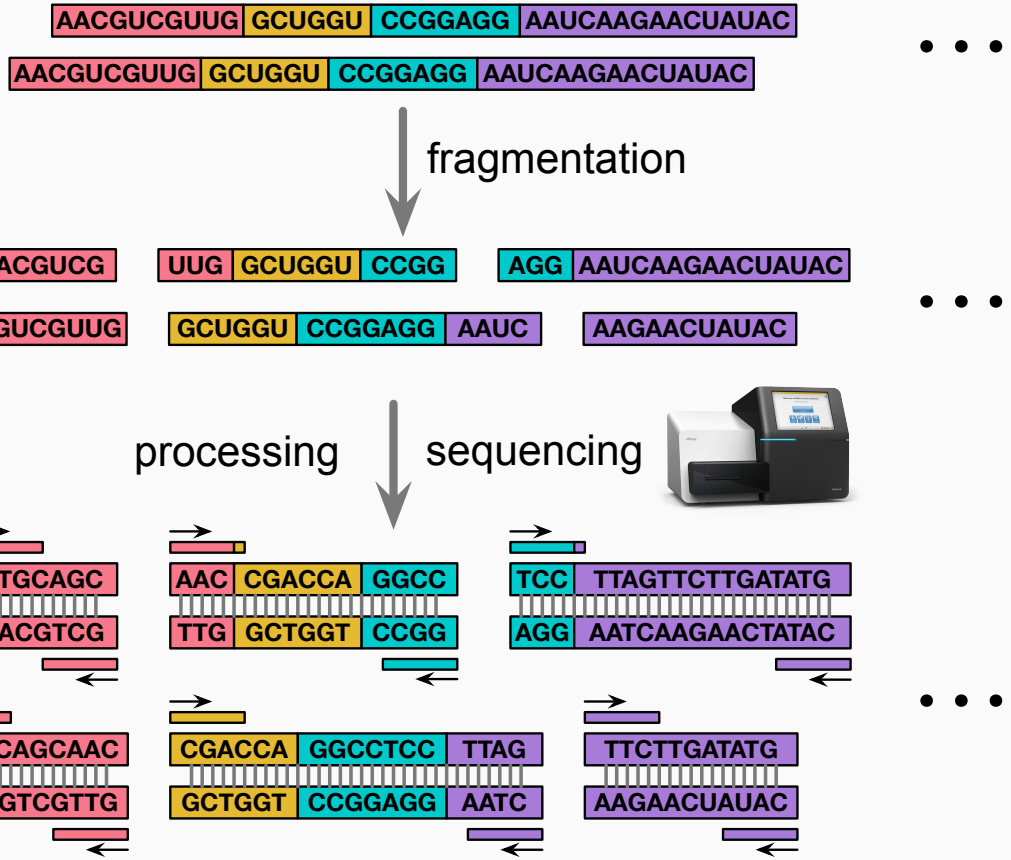

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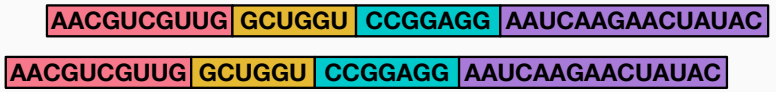

fragmentation

RNA fragments
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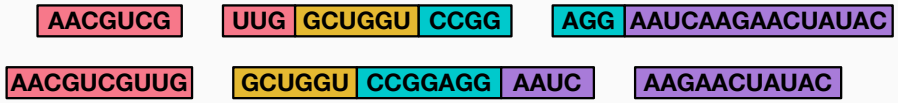
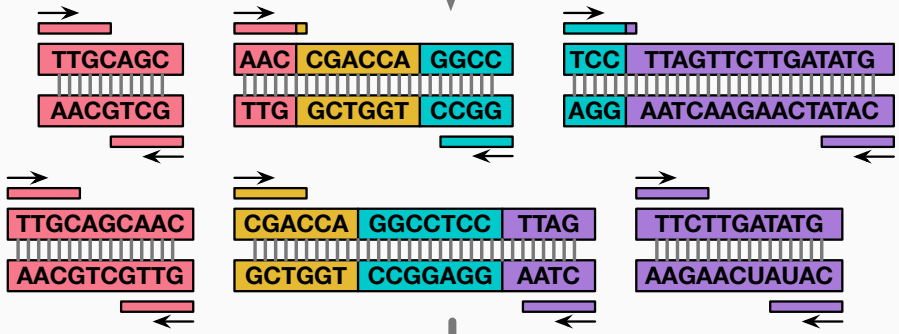
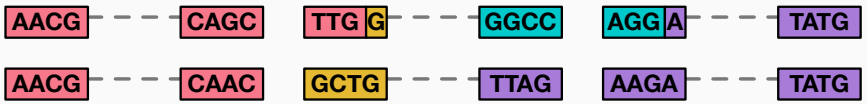


processing sequencing



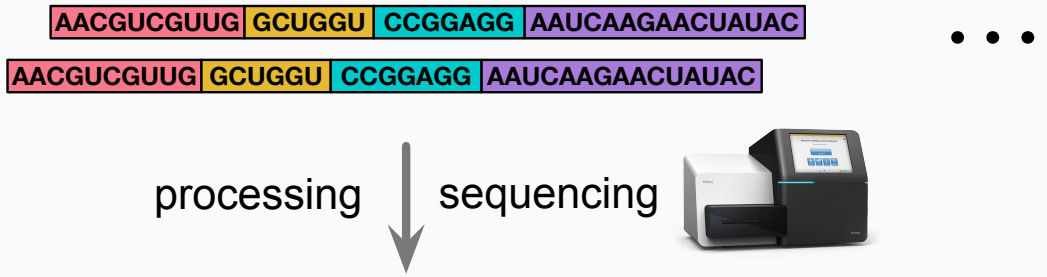
RNA-seq reads
(< 300 bp)



# of RNA-seq reads $\propto$ isoform abundance $\times$ isoform length

Mapping RNA-seq reads to the reference genome

full length RNA isoforms
(1712 bp on average)



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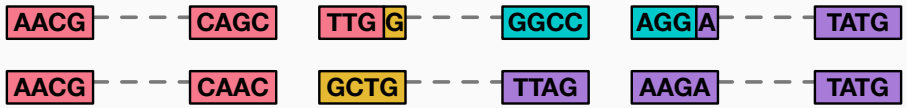
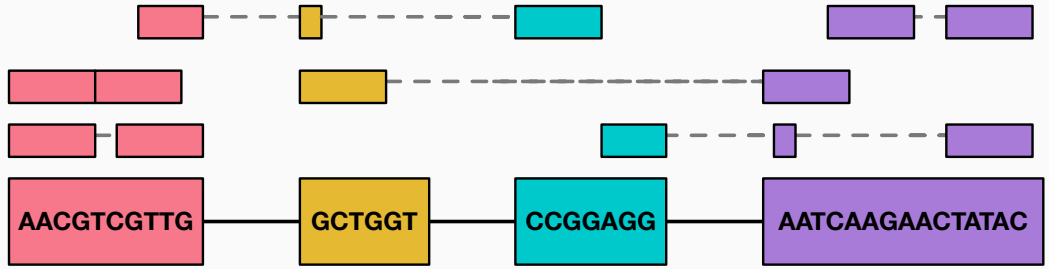
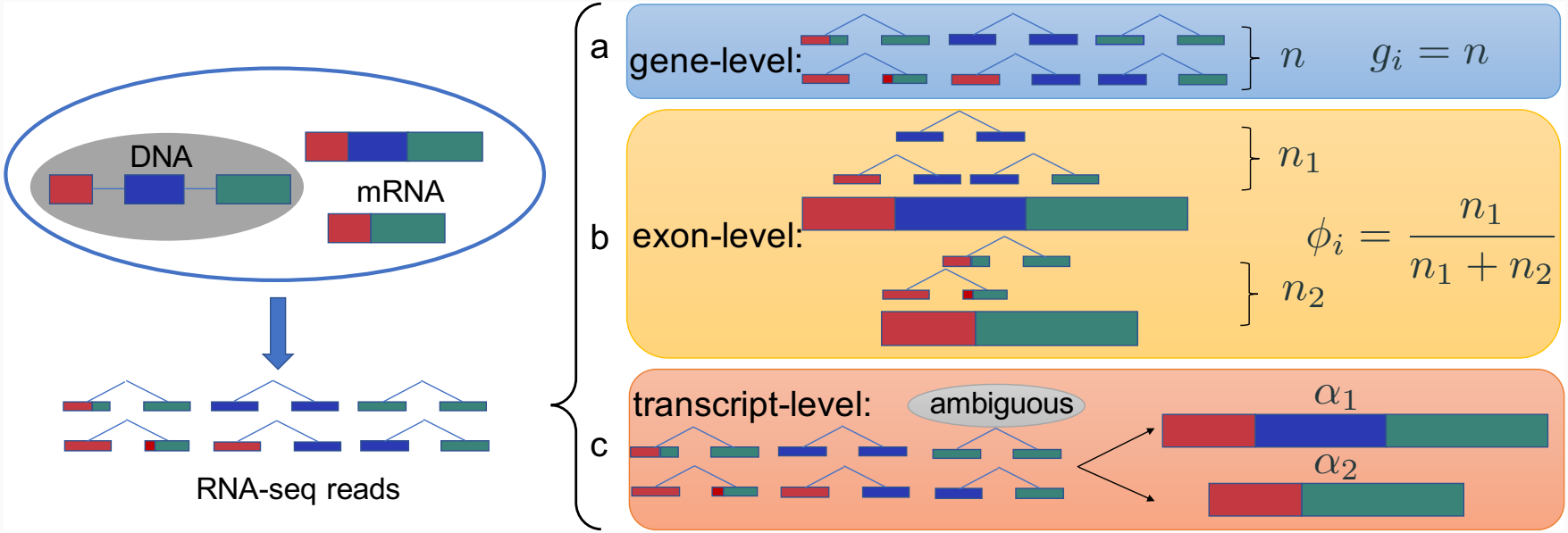


RNA-seq reads
aligned to genome

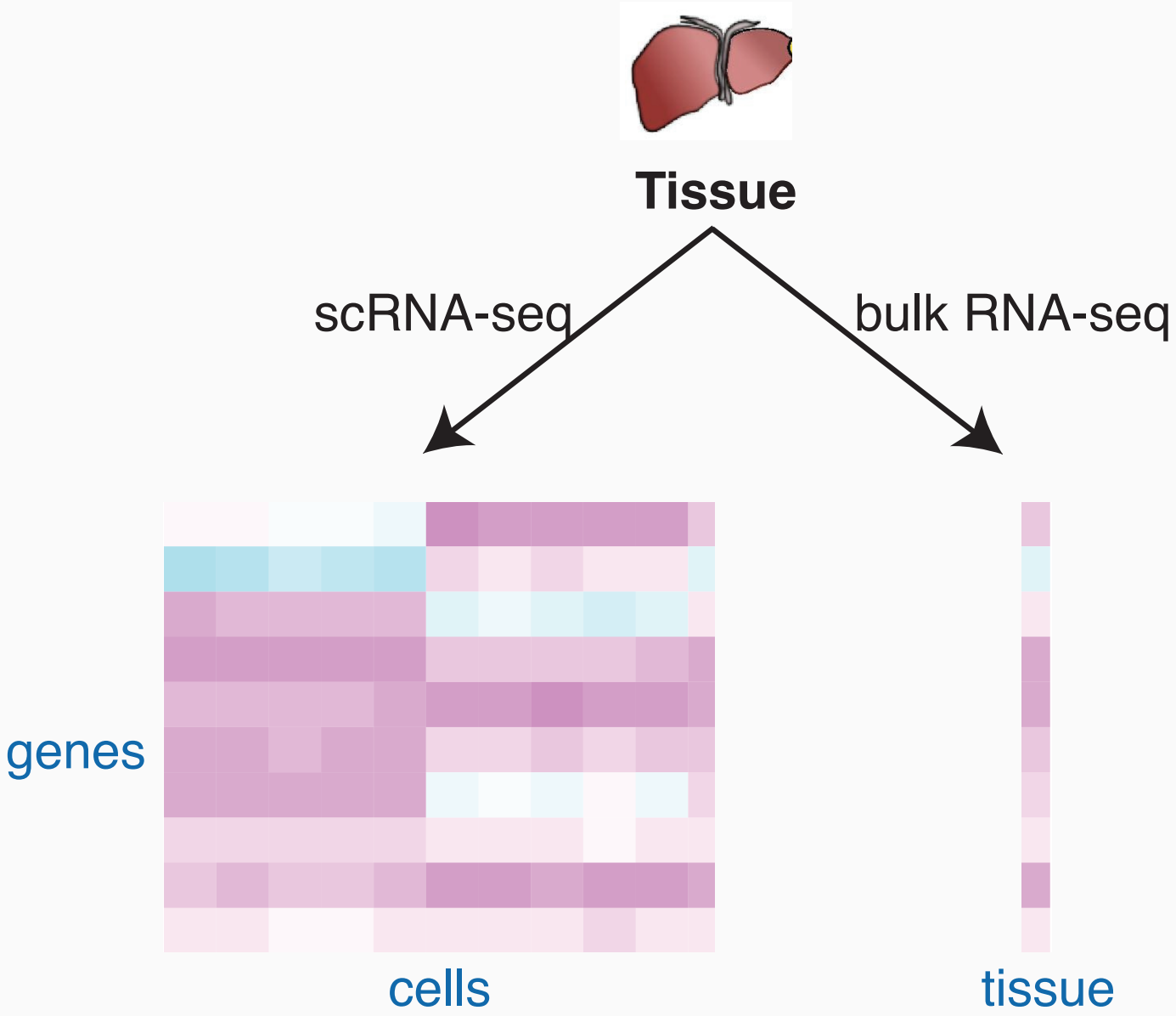


Reference-based RNA-seq data analysis

1. Align RNA-seq reads to a reference genome
2. Analyze aligned reads at three levels

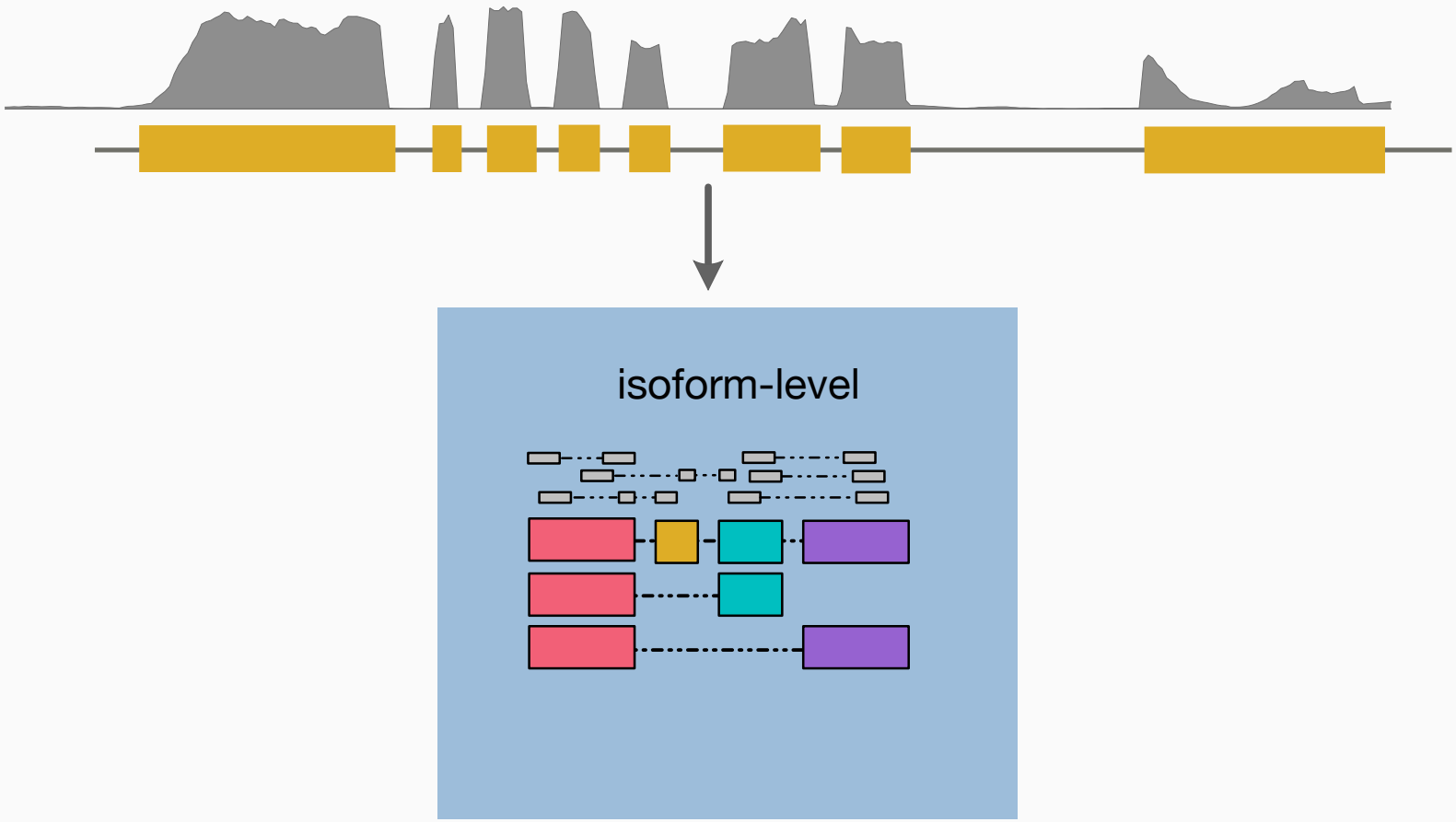


Single-cell (sc) vs. bulk RNA-seq at the gene level



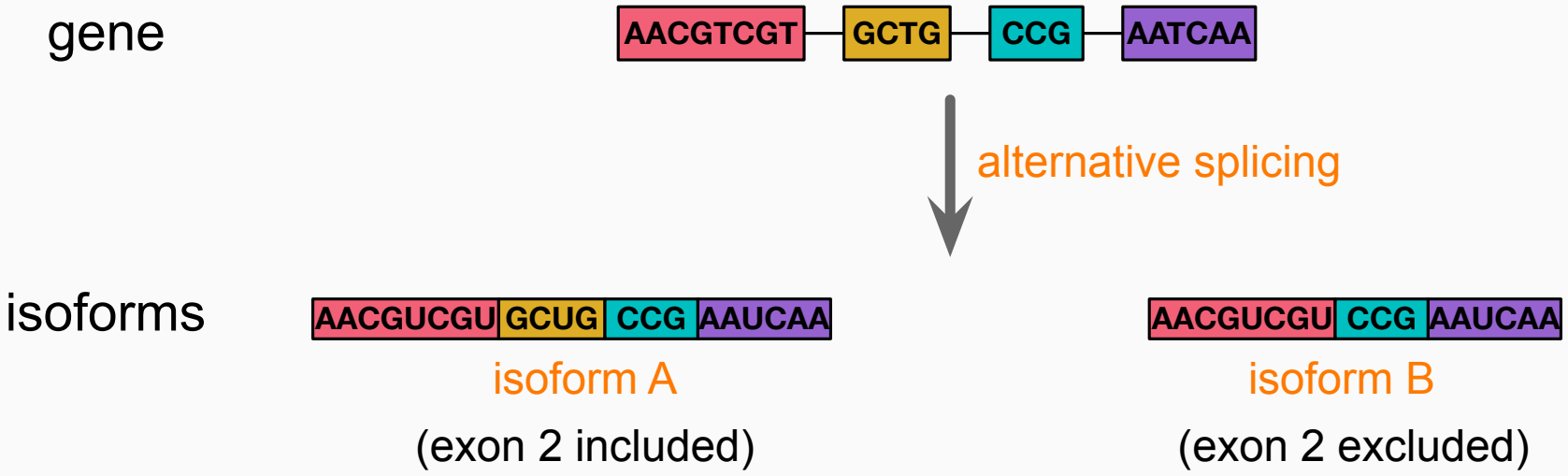
Bulk RNA-seq: transcript/isoform discovery & quantification

AIDE: annotation-assisted isoform discovery



Isoform discovery: which isoforms are expressed?

- More than 90% genes undergo alternative splicing in mammals [Hooper, *Human Genomics*, 2014].
- At least 35% genetic diseases involve abnormal splicing [Manning et al., *Nature Reviews Mol. Cell Biol.* 2017].



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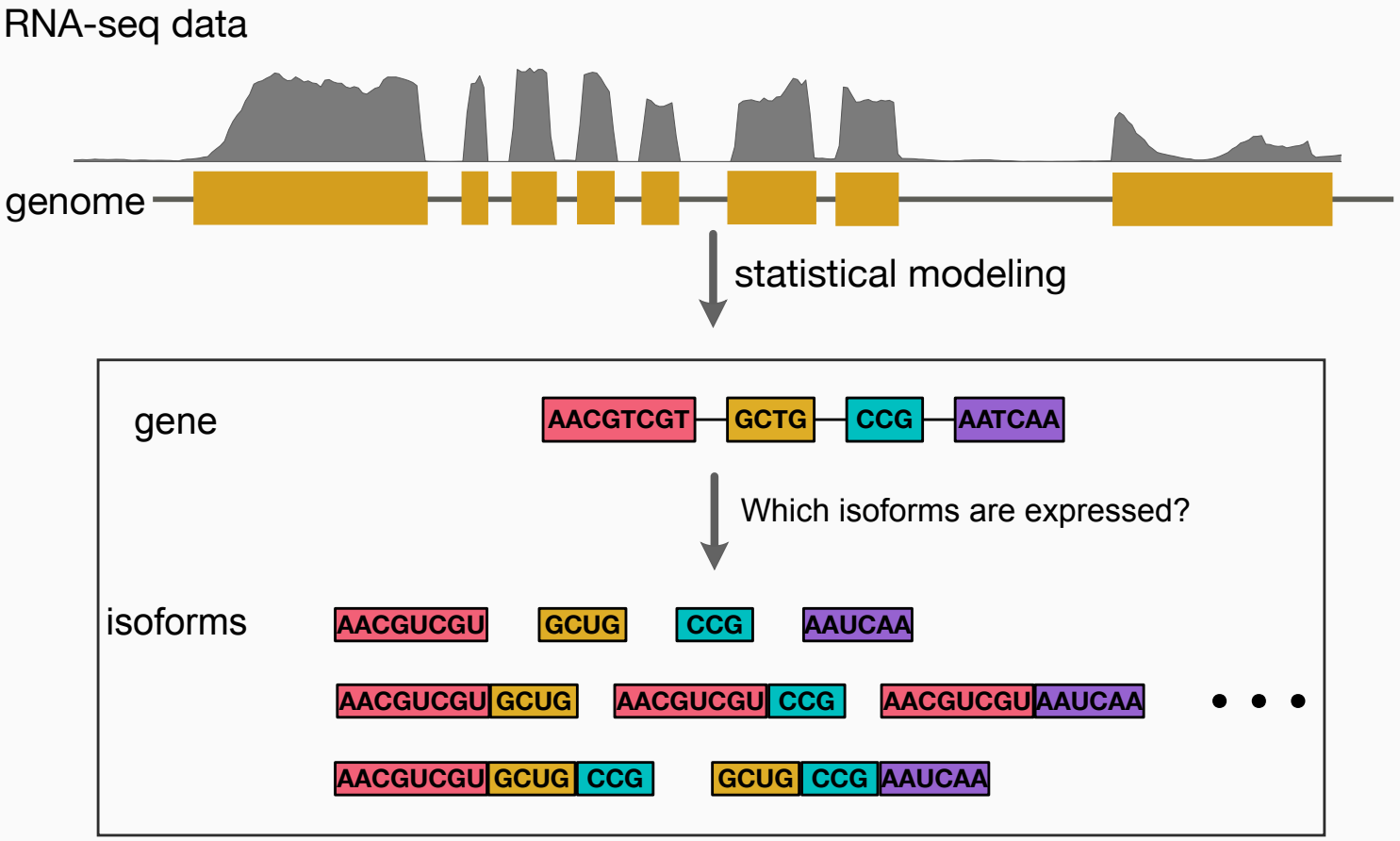
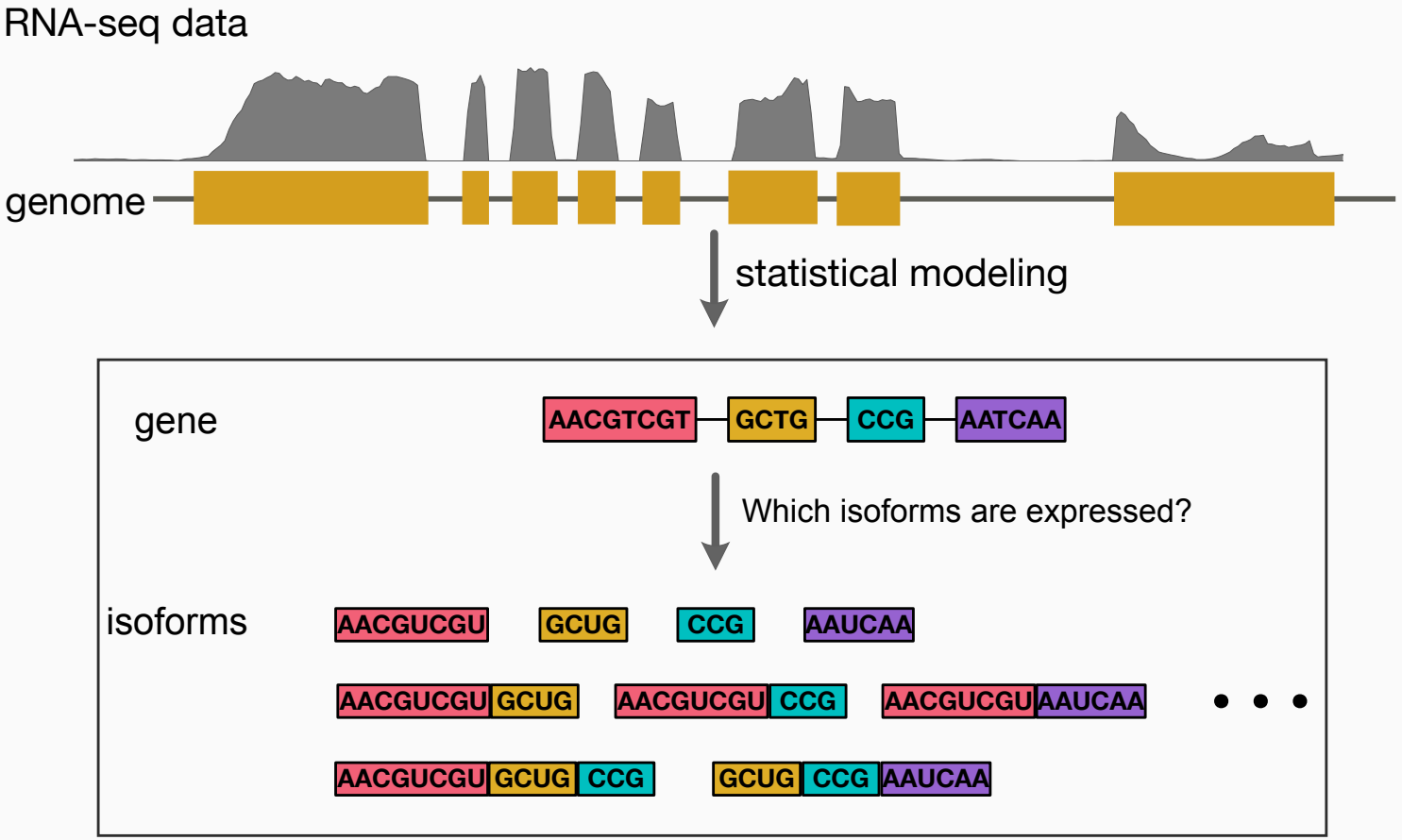


Challenge 1: large number of candidate isoforms

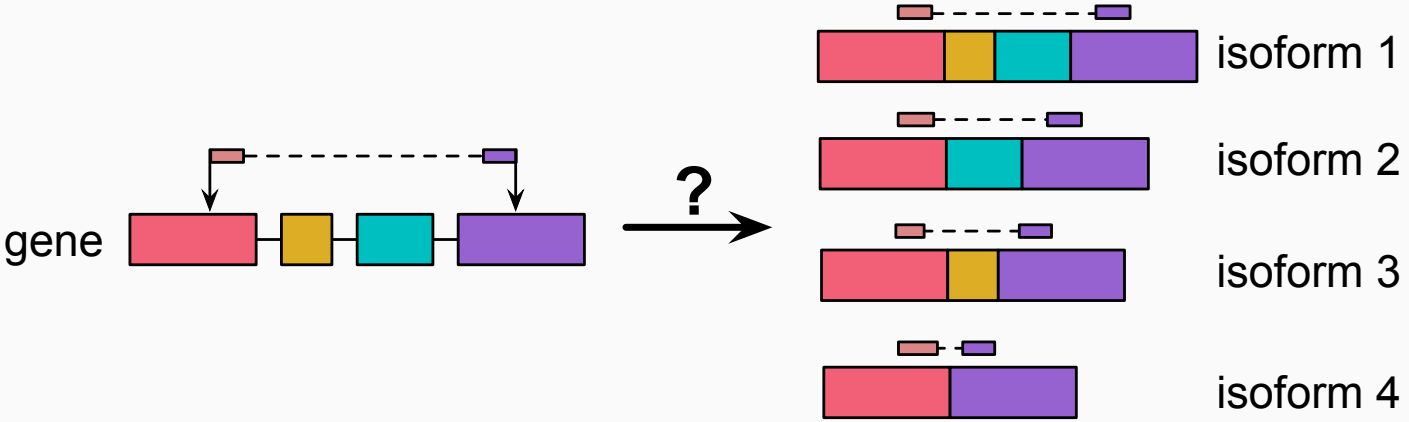
Variable size (# of candidate isoforms) $= 2^{\#\text{ of exons}} - 1$



For this 4-exon gene, $2^4 - 1 = 15$ candidate isoforms

Challenge 2: great information loss

- RNA-seq reads are very short compared with full-length isoforms.
- Most RNA-seq reads do not uniquely map to a single isoform.



- Technical biases introduced into RNA-seq experiments.

Existing isoform discovery methods

State-of-the-art methods for isoform discovery:

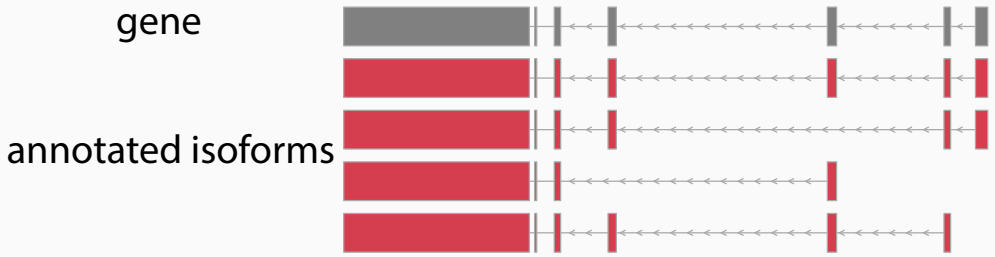
- SIIER [Jiang et al., *Bioinformatics*, 2009]
- Cufflinks [Trapnell et al., *Nature Biotechnology*, 2010]
- SLIDE [Li et al., *Proc. Natl. Acad. Sci.* 2011]
- StringTie [Pertea et al., *Nature Biotechnology*, 2015]
- ...

Limitations:

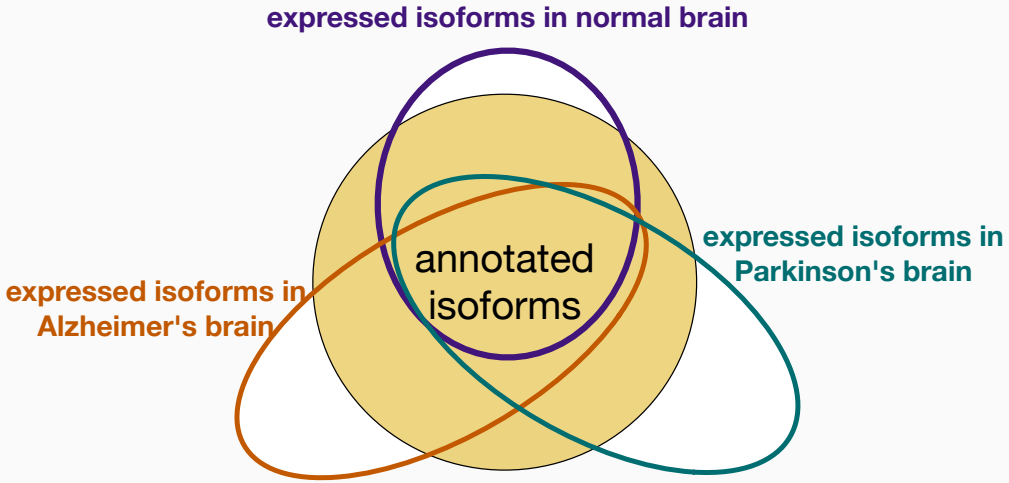
1. Low accuracy for genes with complex splicing structures.
2. Difficult to improve isoform-level performance.
   [Kanitz et al., *Genome Biology*, 2015]
3. Usage of annotations results in false positives.

Usage of annotations results in false positives

Annotated isoforms are experimentally validated:



- *Ensembl* database: $203,903$ isoforms
  [Zerbino et al., *Nucleic Acids Research*, 2017]



False positives → false discoveries

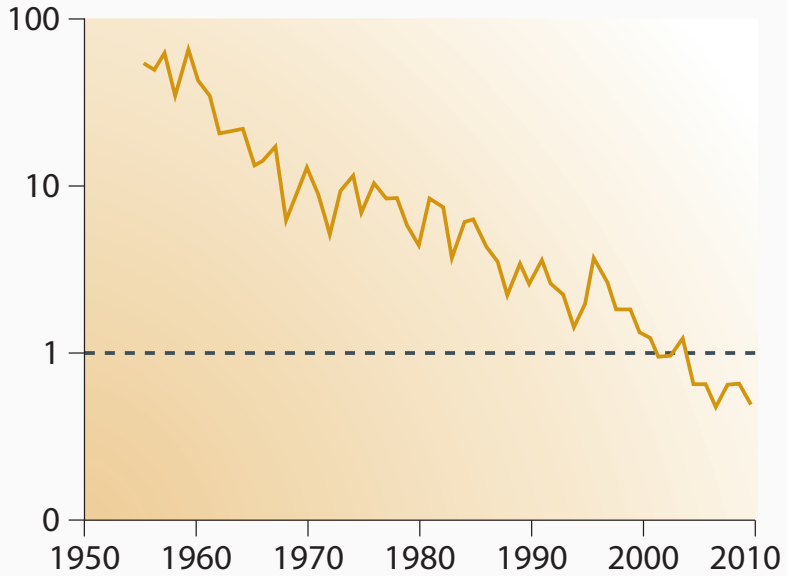
Essay

Why Most Published Research Findings Are False

John P. A. Ioannidis

68,836 Save	3,644 Citation
2,622,757 View	10,479 Share

Number of drugs per billion US$ R&D spending



[Scannell et al., *Nat. Rev. Drug Discov.* 2012]

Highlights of the AIDE method

1. Selectively leverage annotation information to increase the precision and robustness of isoform discovery.

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3. Conservatively identify isoforms that make statistically significant contributions to explaining the observed RNA-seq reads.

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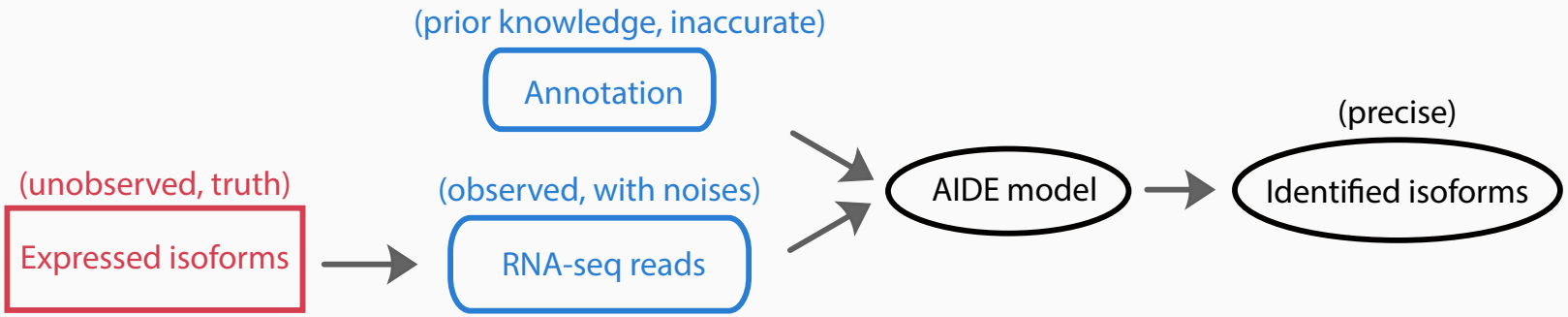
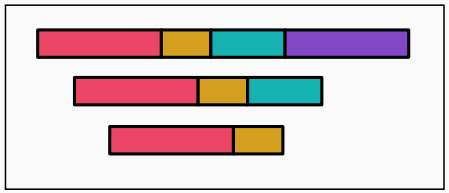
2. Practical probabilistic model to account for technical biases.
3. Conservatively identify isoforms that make statistically significant contributions to explaining the observed RNA-seq reads.
4. First method to control false discoveries by employing a statistical testing procedure.

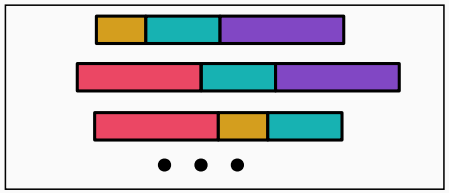


The stepwise selection in AIDE: two stages

annotated isoforms:



non-annotated isoforms:



Stage 1: candidates are annotated isoforms only

Initialization → Forward step ⇄ Backward step

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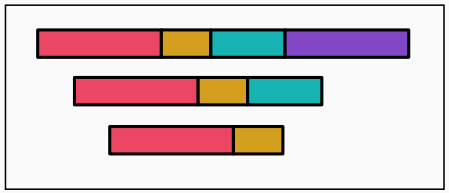

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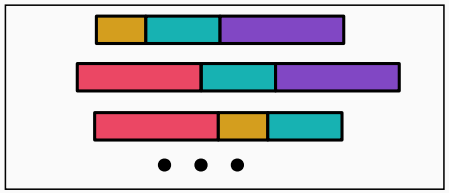

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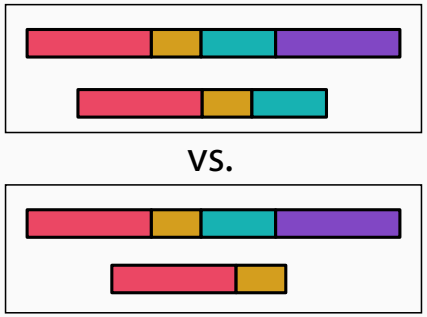

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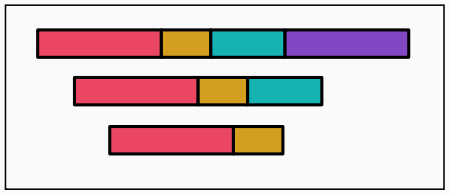

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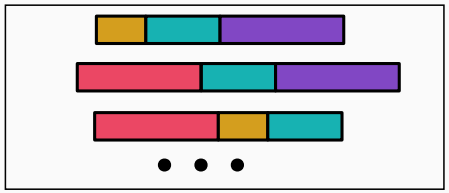

Stage 1: candidates are annotated isoforms only

Initialization → Forward step → Backward step

selected based on MLE

The stepwise selection in AIDE: two stages

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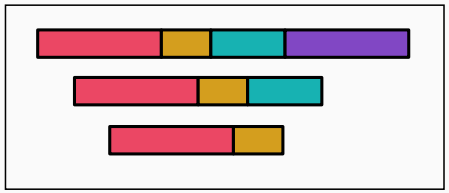

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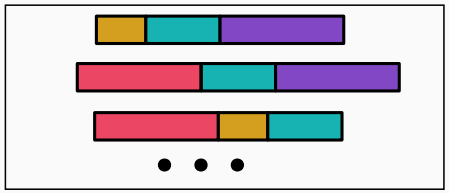

Stage 1: candidates are annotated isoforms only

Initialization → Forward step Backward step

vs.
LRT

The stepwise selection in AIDE: two stages

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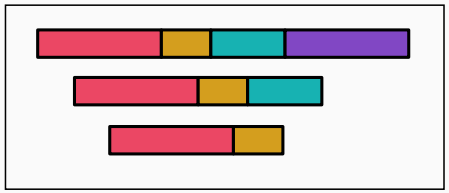

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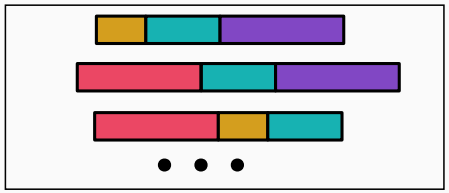

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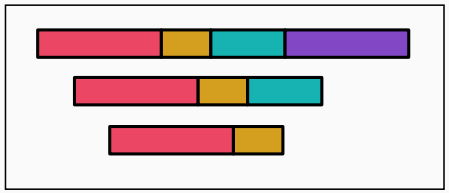

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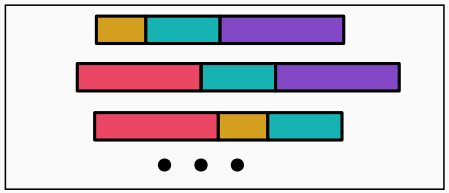

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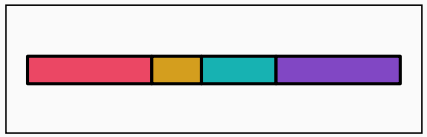
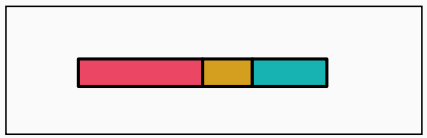


vs.



The stepwise selection in AIDE: two stages

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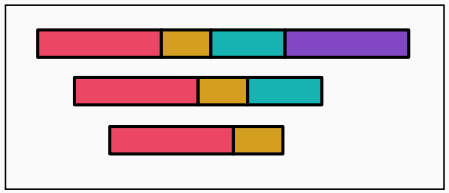

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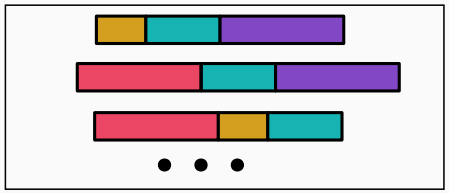

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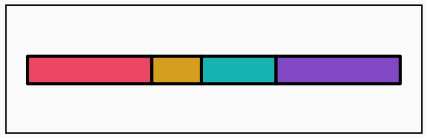

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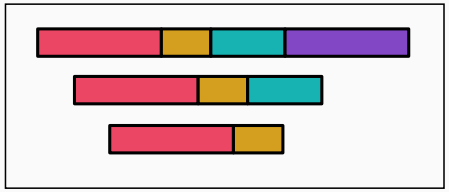

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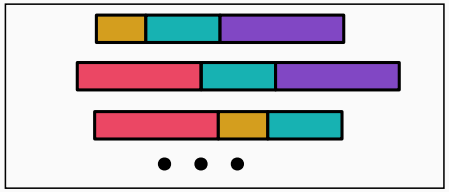

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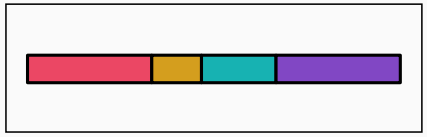
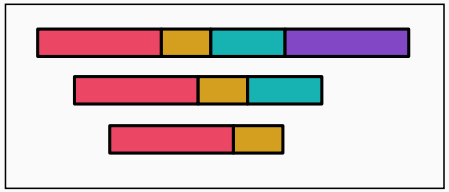


The stepwise selection in AIDE: two stages

annotated isoforms:



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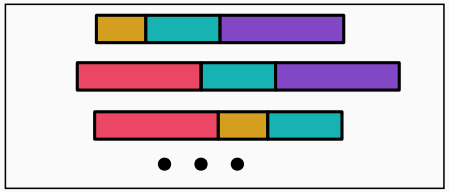

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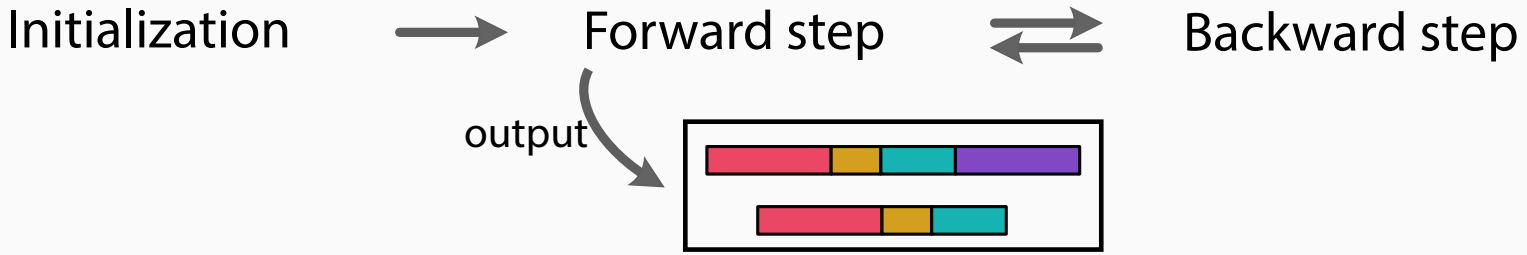

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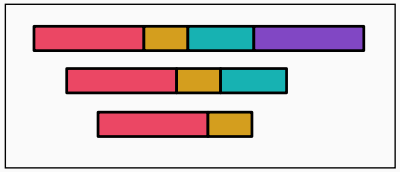

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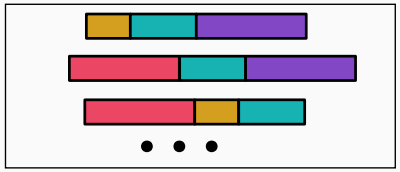


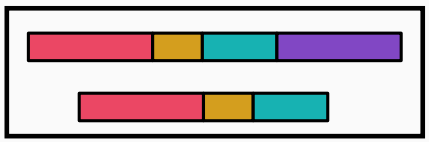
Stage 1: candidates are annotated isoforms only

Initialization → Forward step ⇄ Backward step

output

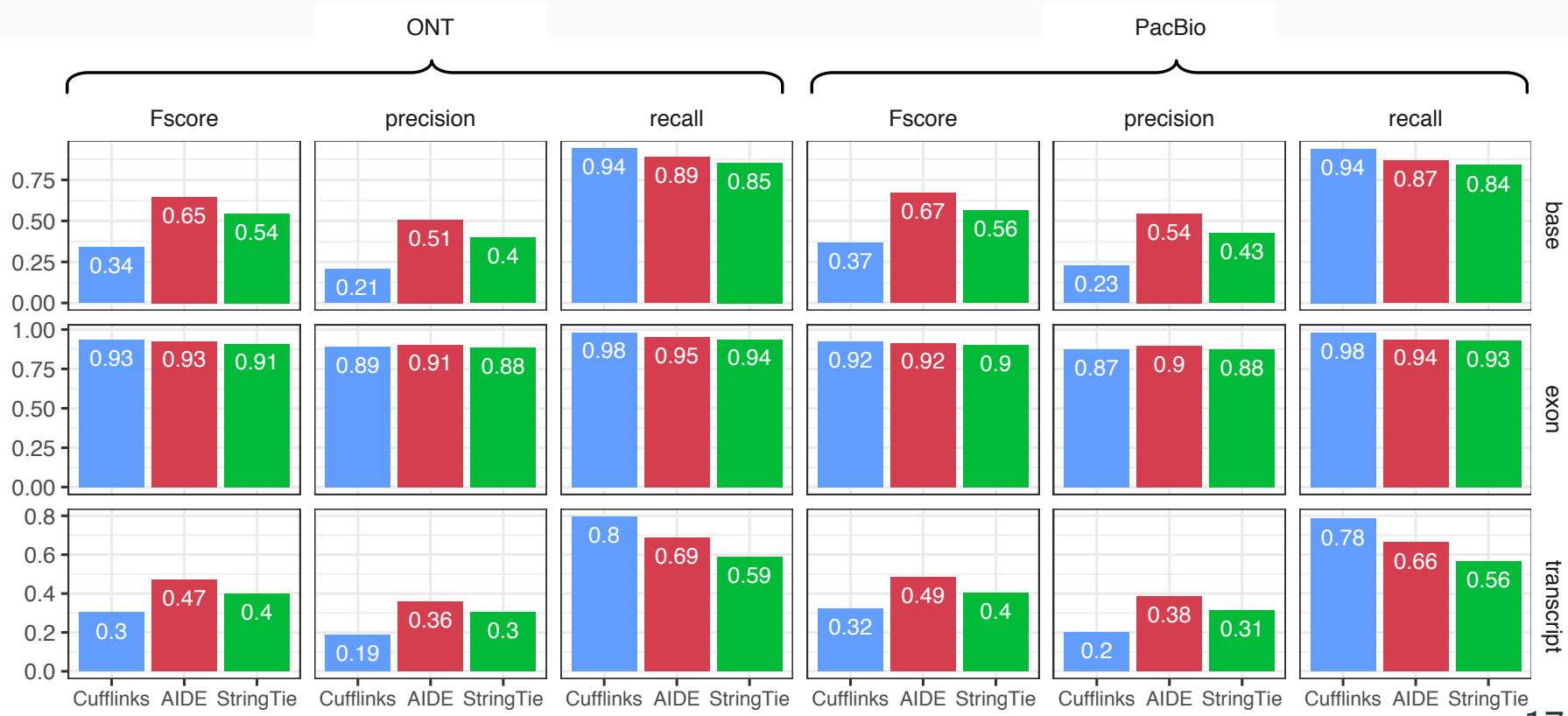
Stage 2: candidates are all possible isoforms

Initialization → Forward step ⇄ Backward step



AIDE outperforms state-of-the-art methods

- Human embryonic stem cells
- Input: Illumina RNA-seq data
- Evaluation: PacBio and Nanopore ONT RNA-seq data



AIDE effectively reduces false discoveries in real data

- Data: breast cancer RNA-seq samples
- Six genes:
  - isoforms identified only by Cufflinks but not by AIDE
  - experimental validation (PCR)

AIDE effectively reduces false discoveries in real data

- Data: breast cancer RNA-seq samples
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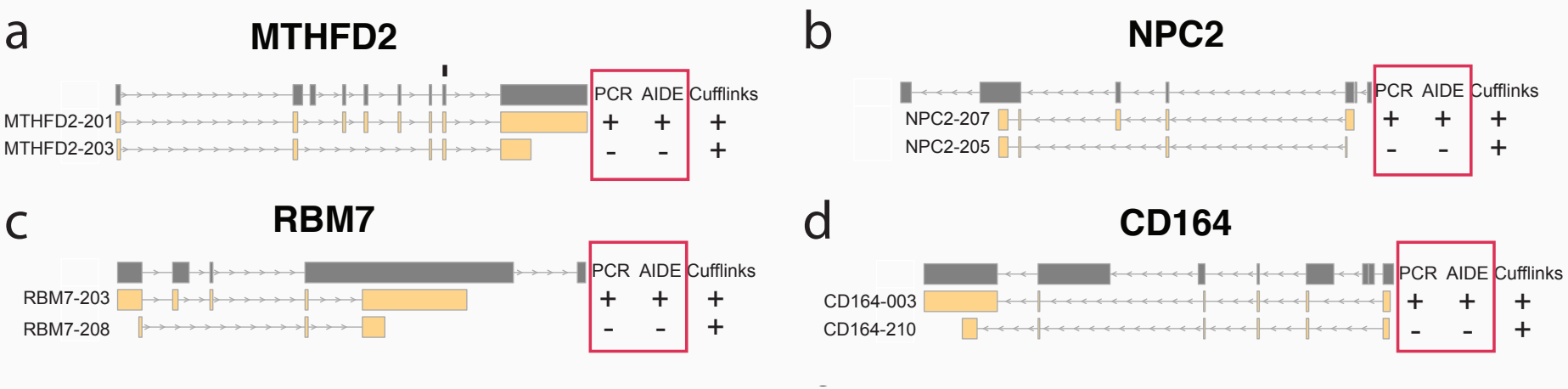
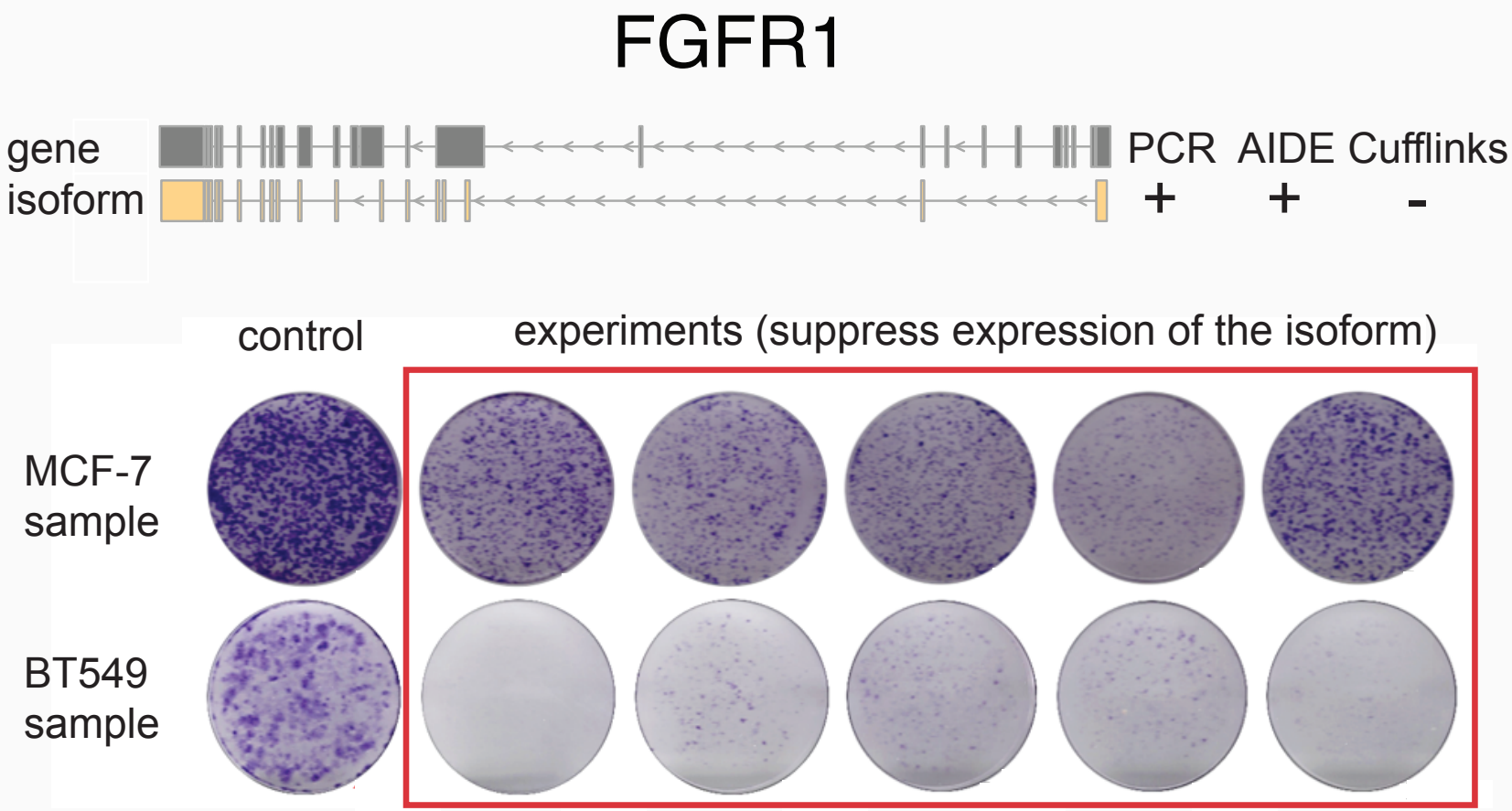
  - experimental validation (PCR)
- Four genes:

  the isoforms uniquely predicted by Cufflinks were false positives



AIDE discovers isoforms with biological significance



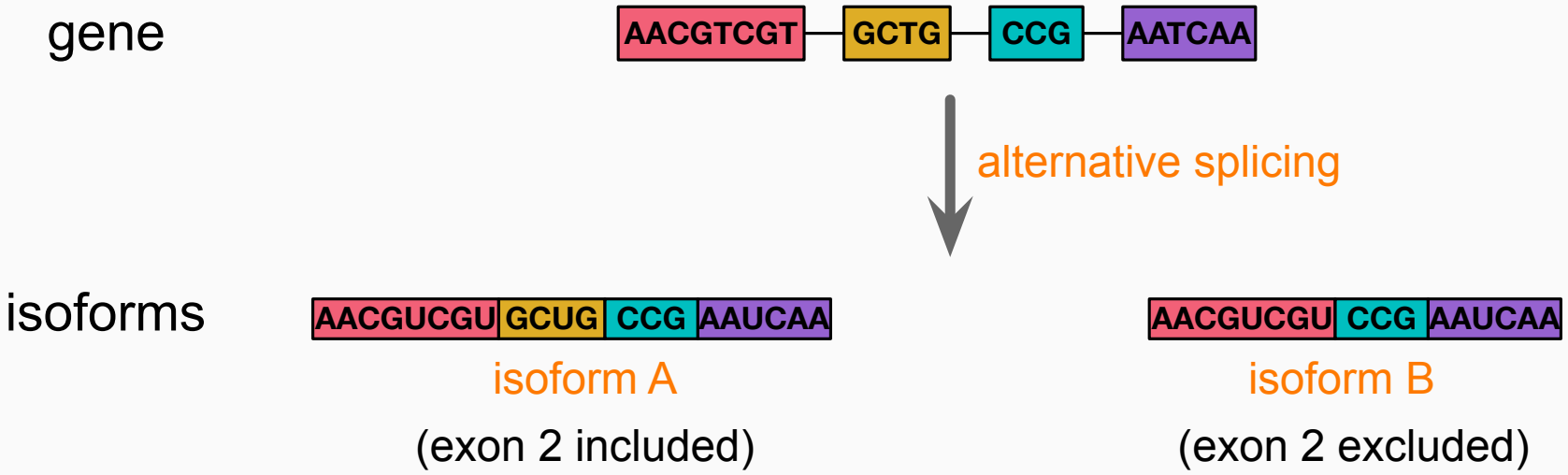
Summary of the AIDE method

- The first isoform discovery method that directly controls false discoveries by implementing the statistical model selection principle.
- Software: `https://github.com/Vivianstats/AIDE`
- Manuscript:

In press at *Genome Research*.

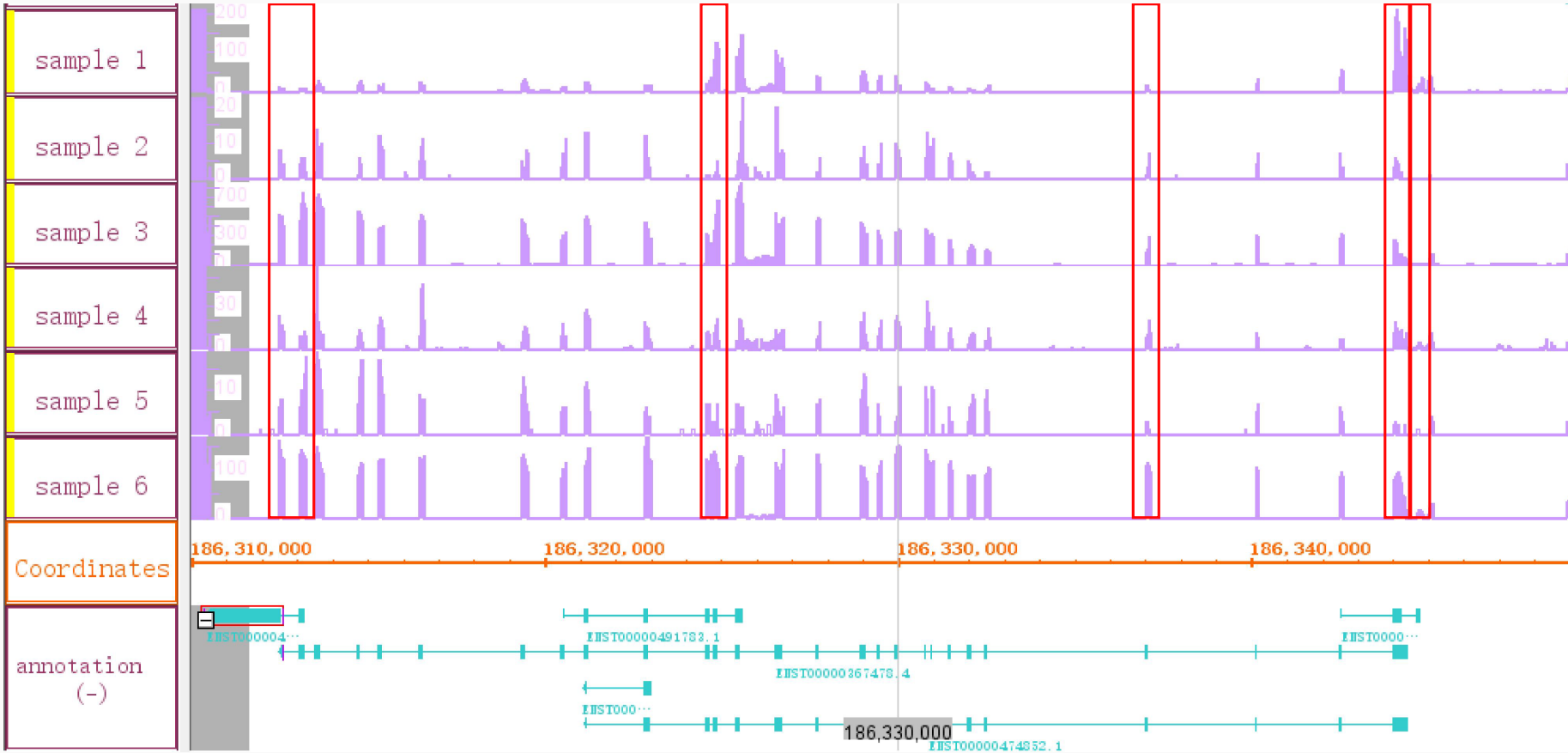
Isoform quantification: what are the isoform expression levels?

- More than 90% genes undergo alternative splicing in mammals [Hooper, *Human Genomics*, 2014].
- At least 35% genetic diseases involve abnormal splicing [Manning et al., *Nature Reviews Mol. Cell Biol.* 2017].



Motivation: multiple human ESC RNA-seq samples

chr1; gene: *TPR*



How to combine multiple RNA-seq samples?

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  - The estimated isoform abundance may be biased by outlier samples

MSIQ

Joint Modeling of **M**ultiple RNA-seq **S**amples for Accurate **I**soform **Q**uantification

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- MSIQ is able to identify a consistent group of samples that are most representative of the biological condition
- MSIQ increases the accuracy of isoform quantification by incorporating the information from multiple samples
- Our proposed hierarchical model is an umbrella framework that are generalizable to incorporate more delicate consideration of read generating mechanisms

Paper and Software

MSIQ: joint modeling of multiple RNA-seq samples for accurate isoform quantification

by Wei Vivian Li, Anqi Zhao, Shihua Zhang, and Jingyi Jessica Li

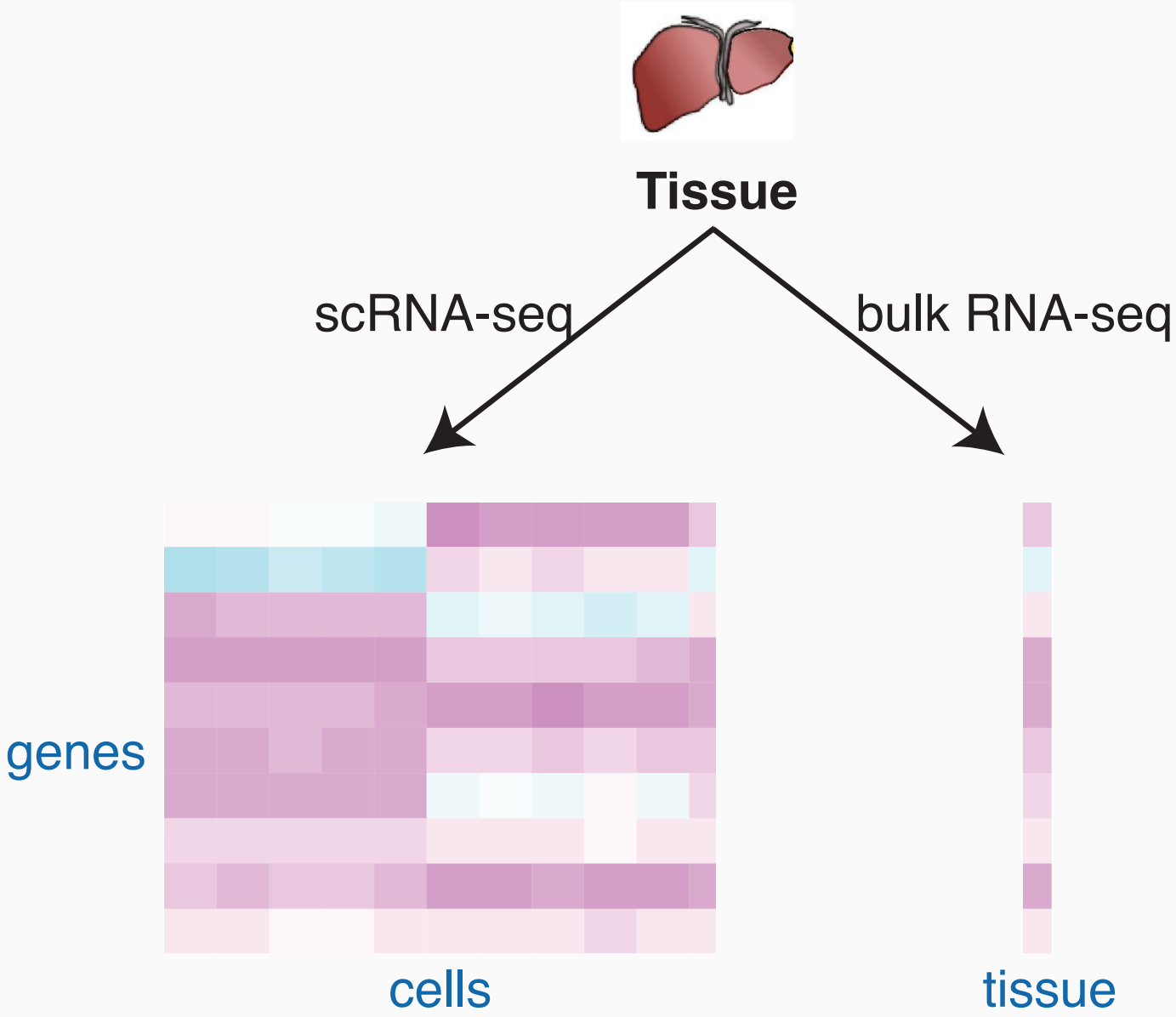
Annals of Applied Statistics 12(1):510–539

R package `MSIQ`

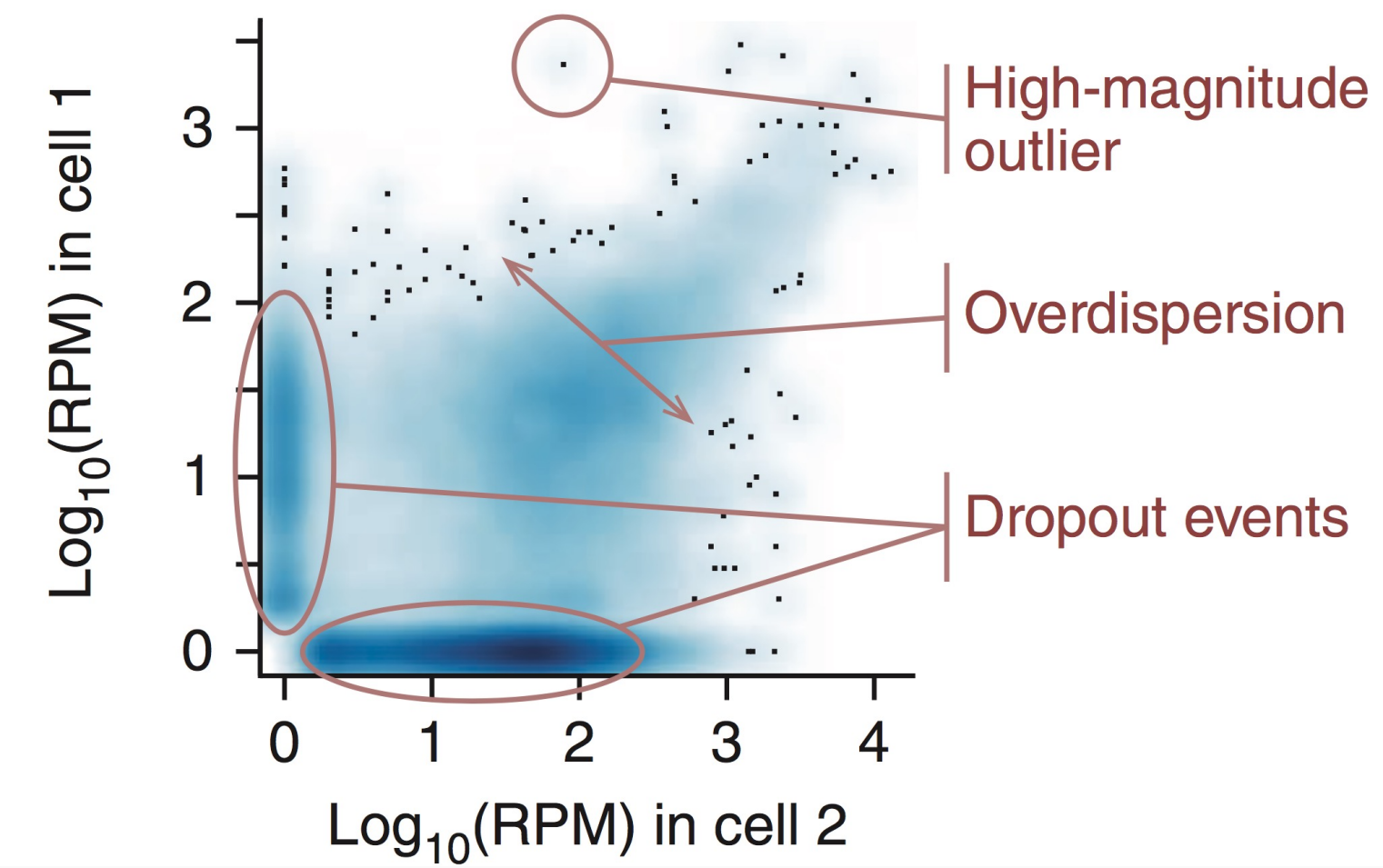
`http://github.com/Vivianstats/MSIQ`

Single-cell RNA-seq: dropout imputation & experimental design

scRNA-seq vs. bulk RNA-seq at the gene level



Dropout events in scRNA-seq



from [Kharchenko et al., *Nature methods*, 2014]

Dropout events in scRNA-seq

- A dropout event occurs when a transcript is expressed in a cell but is entirely undetected in its mRNA profile
- Dropout events occur due to low amounts of mRNA in individual cells
- The frequency of dropout events depends on scRNA-seq protocols
  - Fluidigm C1 platform: ~ 100 cells, ~ 1 million reads per cell
  - Droplet microfluidics: $\sim 10,000$ cells, ~ 100K reads per cell [Zilionis et al., *Nature Protocols*, 2017]
- **Trade-off**: given the same budget, more cells, more dropouts

Genome-wide explicit imputation for dropouts

Why do we need genome-wide explicit imputation methods?

Downstream analyses relying on the accuracy of gene expression measurements:

- differential gene expression analysis
- identification of cell-type-specific genes
- reconstruction of differentiation trajectory

It is important to adjust/correct the false zero expression values due to dropouts

Genome-wide explicit imputation for dropouts

Key points to consider:

- It is not ideal to impute all gene expressions
  - imputing expressions unaffected by dropout would introduce new bias
  - could also eliminate meaningful biological variation
- It is inappropriate to treat all zero expressions as missing values
  - some zero expressions may reflect true biological non-expression
  - zero expressions can be resulted from gene expression stochasticity

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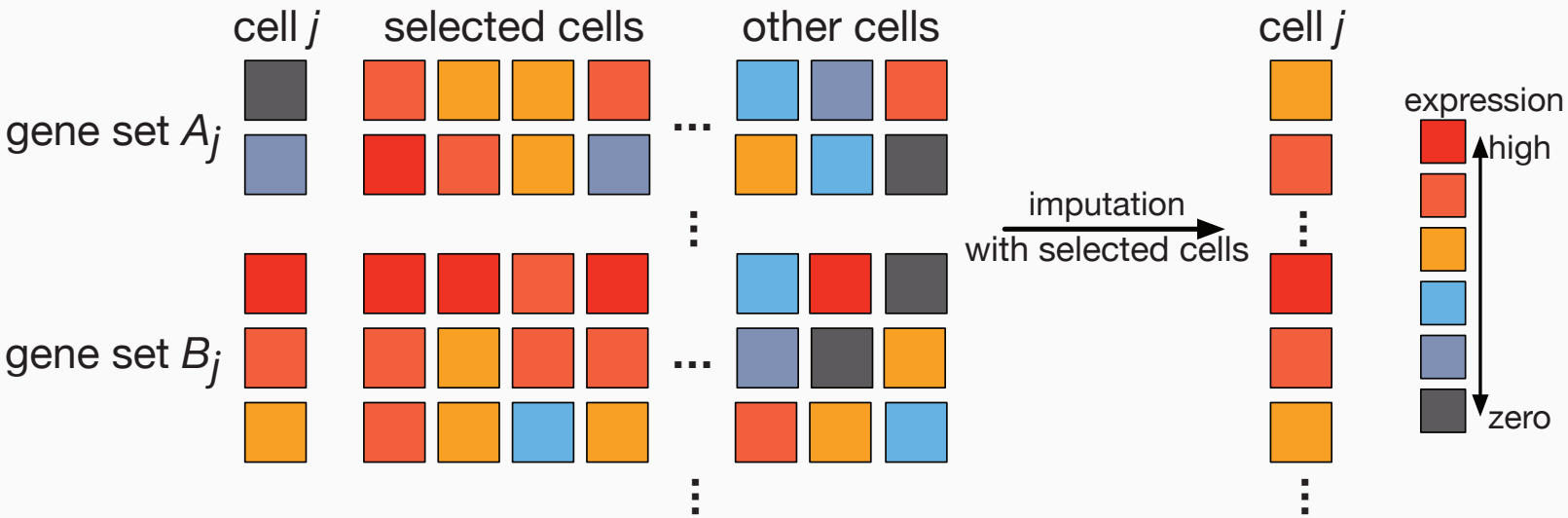
How to determine which values are affected by the dropout events?

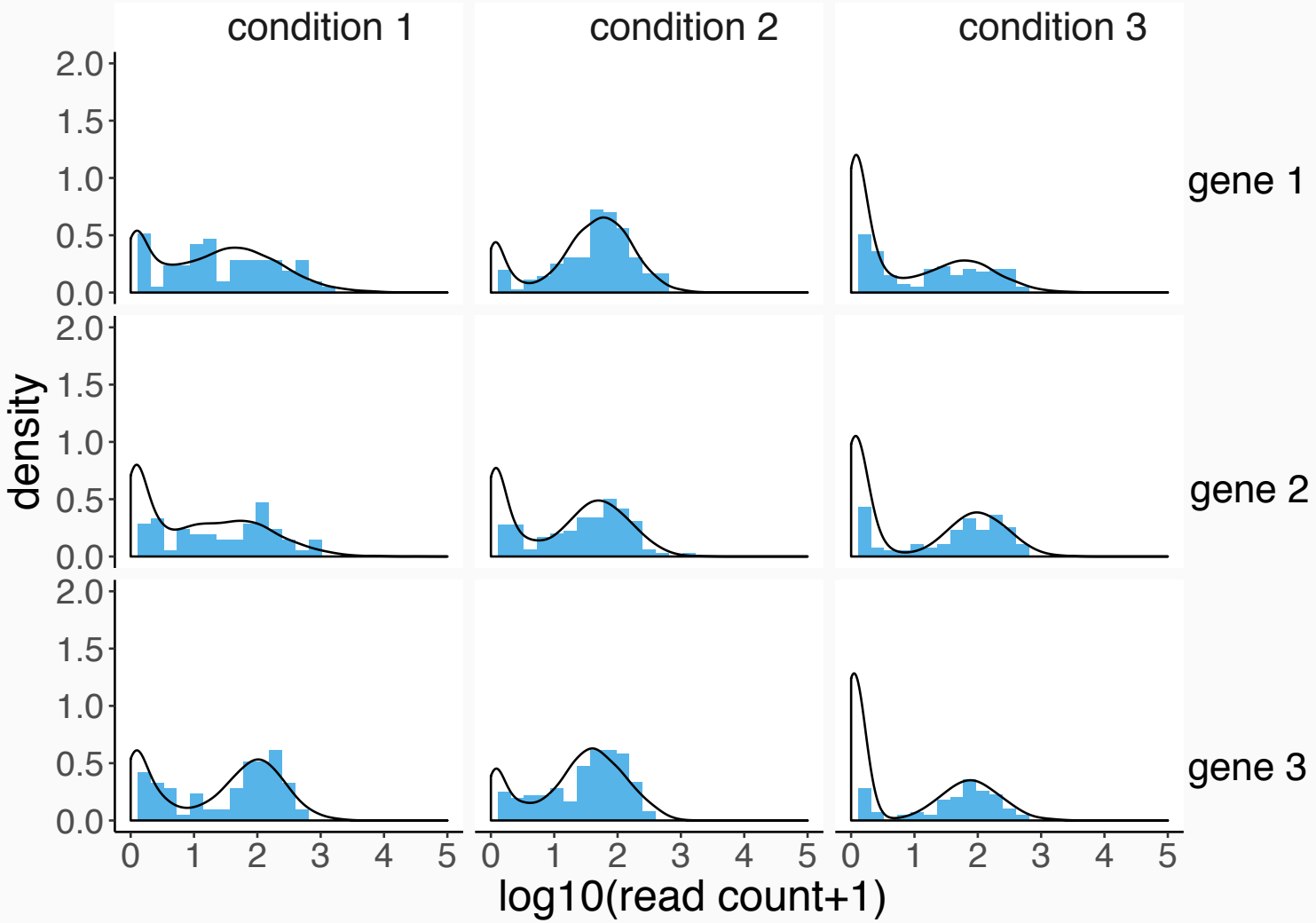
Our method: scImpute

1. For each gene, to determine which expression values are most likely affected by dropout events
2. For each cell, to impute the highly likely dropout values by borrowing information from the same genes' expression in similar cells



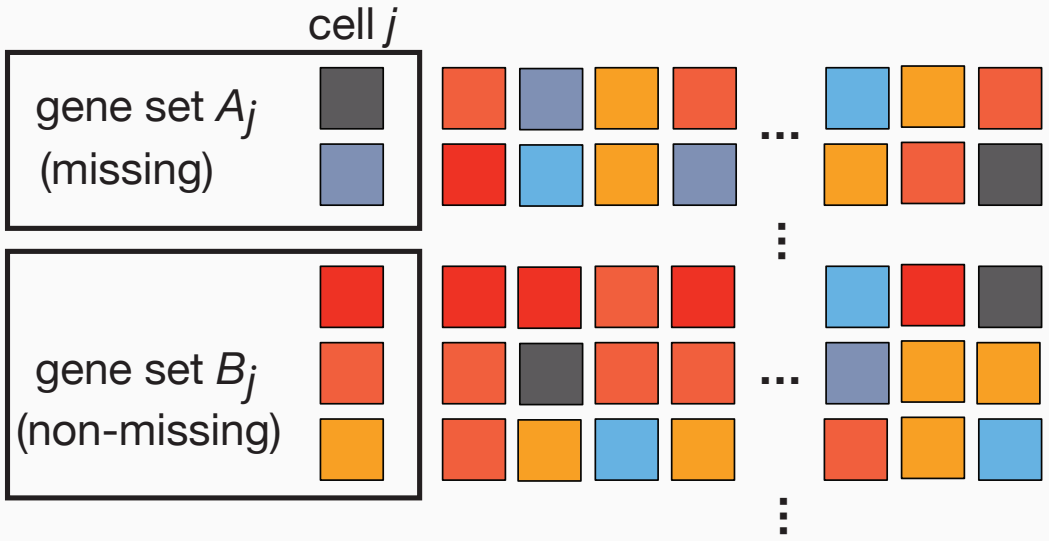
scImpute steps

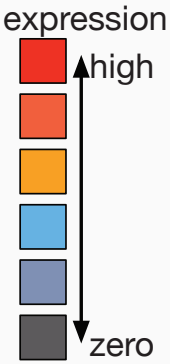
1. Detection of cell subpopulations and outlier cells
2. Identification of dropout values



scImpute steps

3. Imputation of gene expression cell by cell

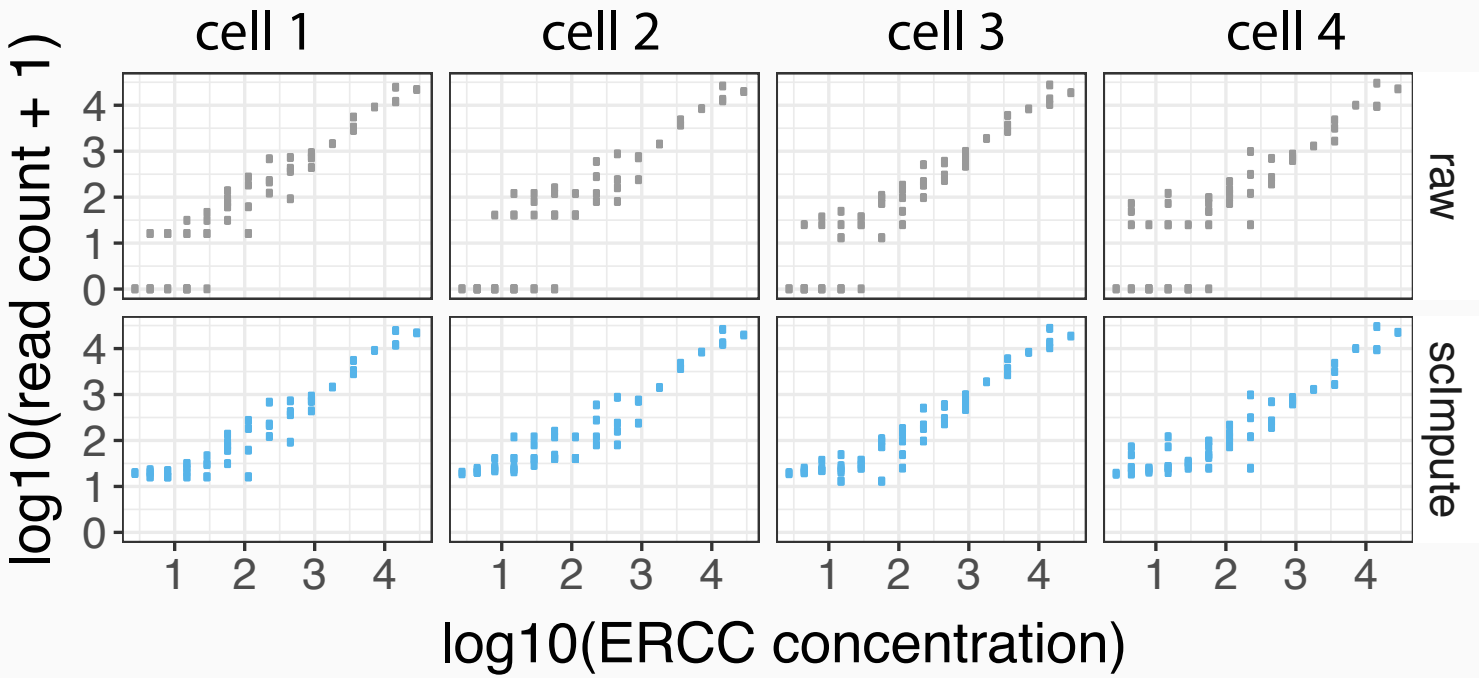




Example 1: ERCC spike-ins

scImpute recovers the true expression of the ERCC spike-in transcripts, especially low abundance transcripts that are impacted by dropout events

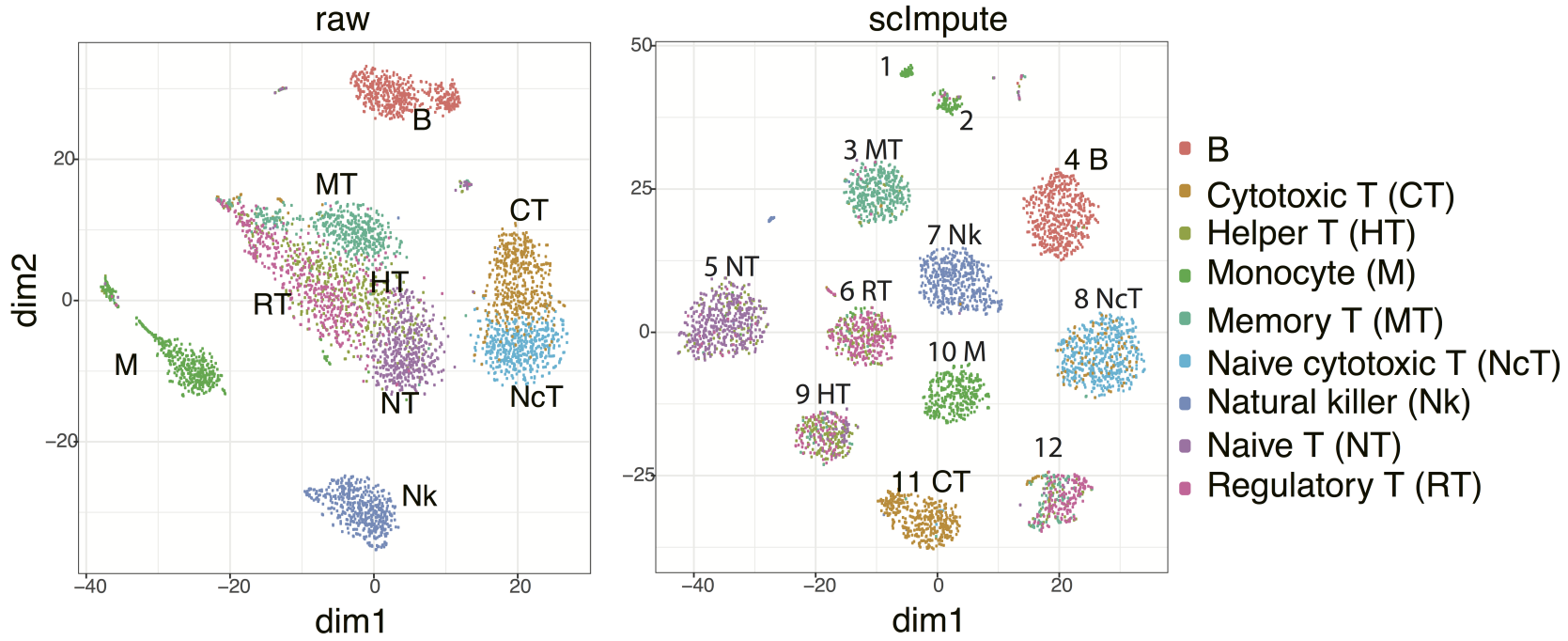
- 3,005 cells from the mouse somatosensory cortex region
- 57 ERCC transcripts



Example 2: cell clustering

4,500 peripheral blood mononuclear cells (PBMCs) from high-throughput droplet-based system 10x genomics [Zheng et al., *Nature communications*, 2017]

Proportion of zero expression is 92.6%



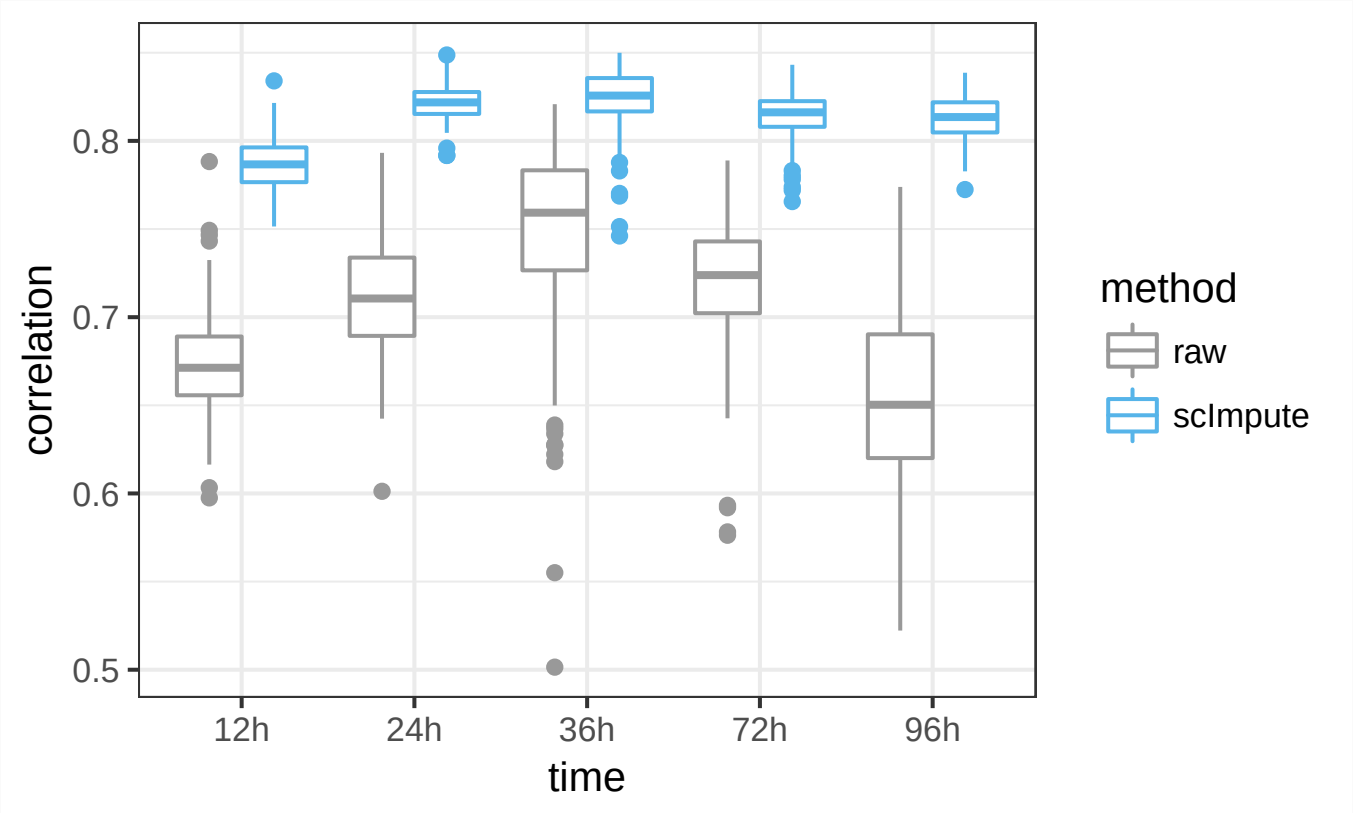
Example 3: gene expression dynamics

Bulk and single-cell time-course RNA-seq data profiled at 0, 12, 24, 36, 72, and 96 h of the differentiation of embryonic stem cells into definitive endorderm cells [Chu et al., *Genome biology*, 2016]

time point	00h	12h	24h	36h	72h	96h	total
scRNA-seq (cells)	92	102	66	172	138	188	758
bulk RNA-seq (replicates)	0	3	3	3	3	3	15

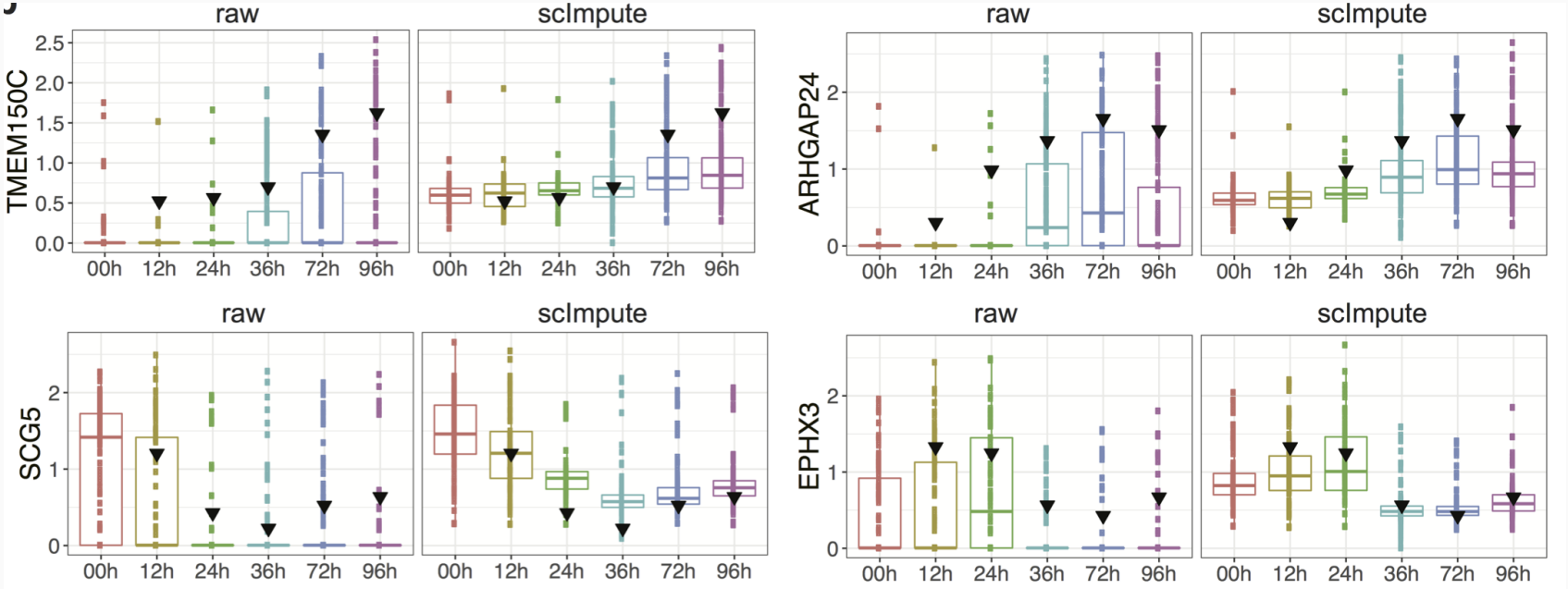
Example 3: gene expression dynamics

Correlation between gene expression in single-cell and bulk data



Example 3: gene expression dynamics

Imputed read counts reflect more accurate gene expression dynamics along the time course



Conclusions

- scImpute is a flexible and easily interpretable statistical method that addresses the dropout events prevalent in scRNA-seq data
- scImpute focuses on imputing the missing expression values of dropout genes, while retaining the expression levels of genes that are largely unaffected by dropout events
- scImpute is compatible with existing pipelines or downstream analysis of scRNA-seq data, such as normalization, differential expression analysis, clustering and classification
- scImpute scales up well when the number of cells increases

Paper and software

An accurate and robust imputation method scImpute for single-cell RNA-seq data

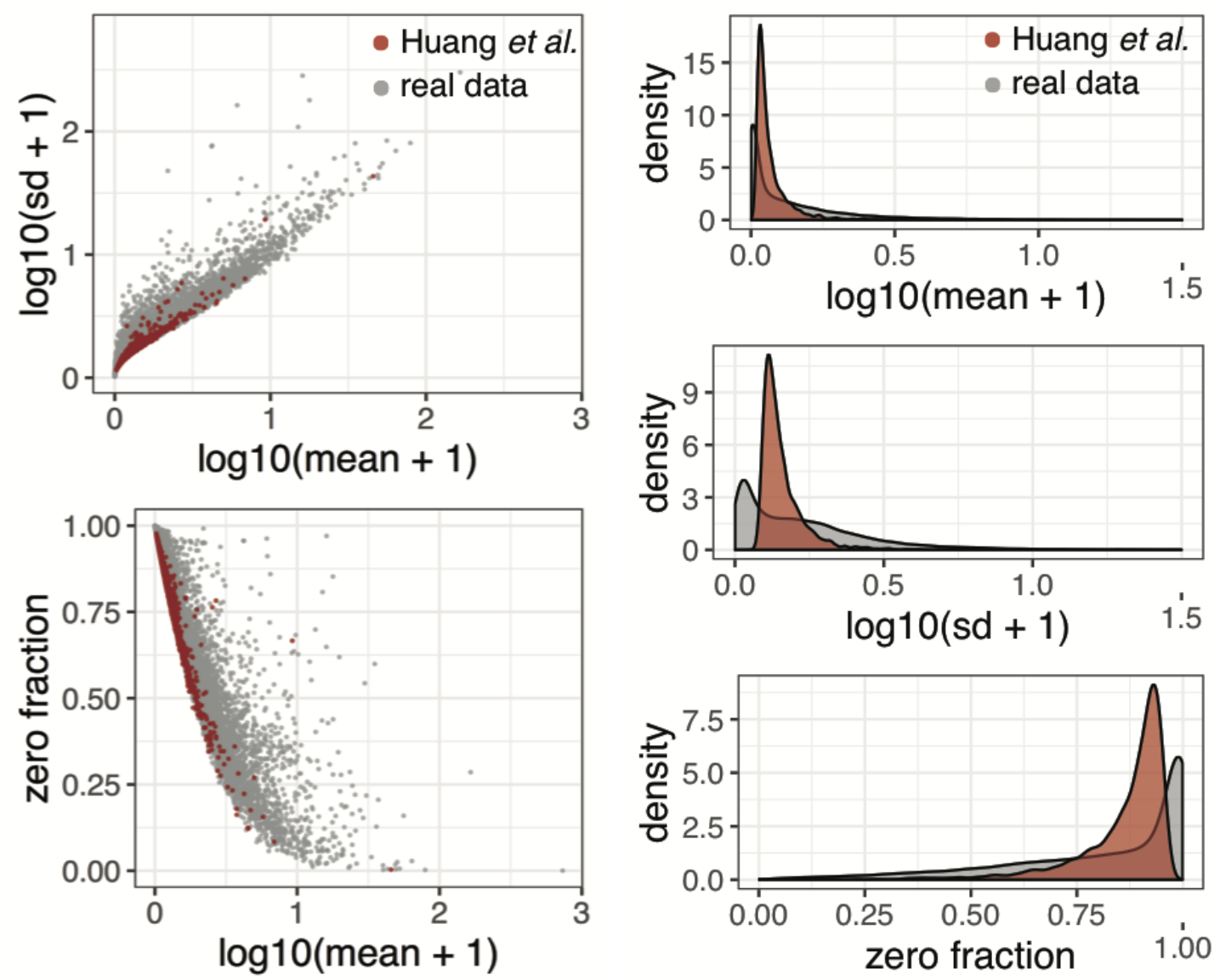
by Wei Vivian Li and Jingyi Jessica Li

Nature Communications 9:997

R package `scImpute`

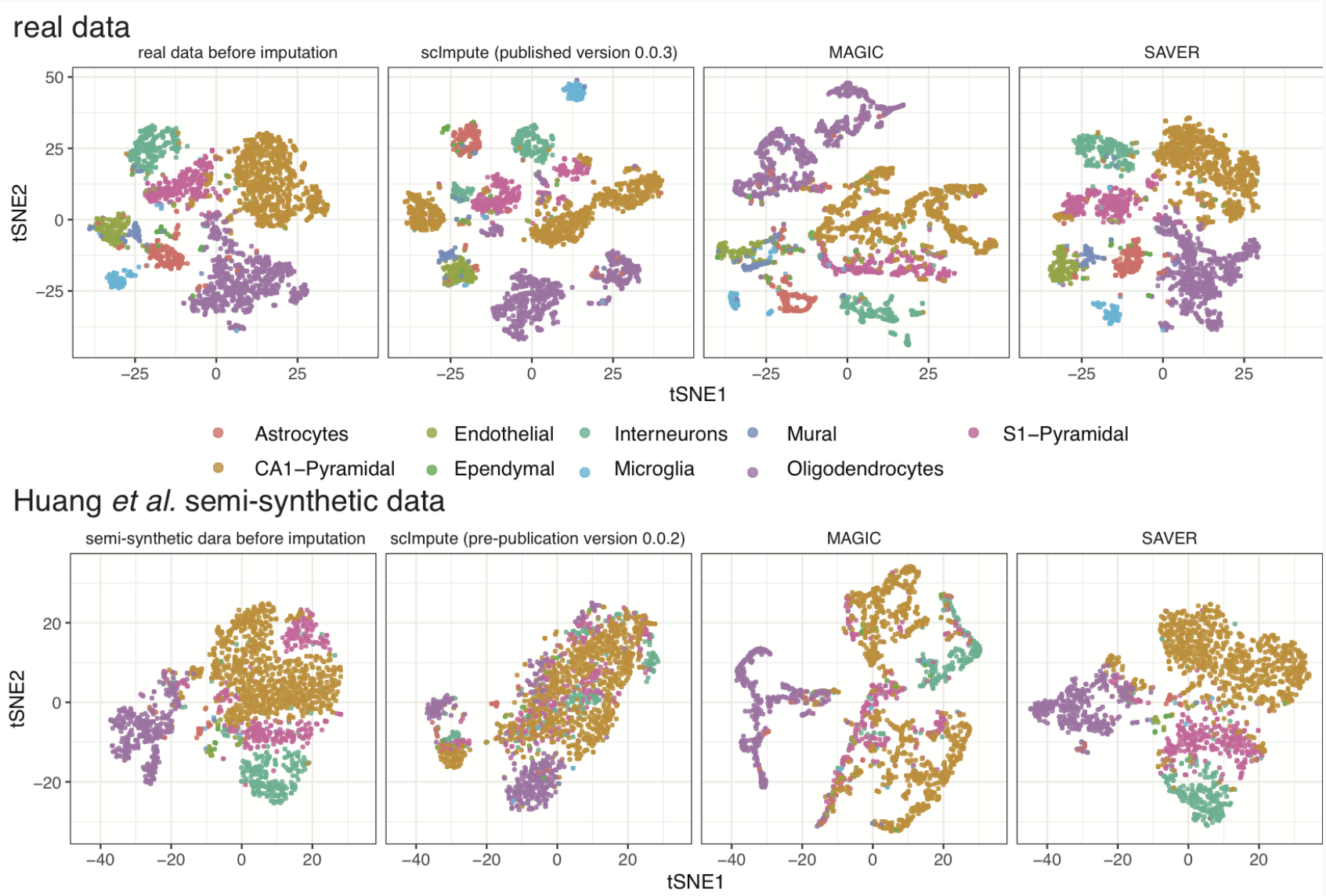
`https://github.com/Vivianstats/scImpute`

Real vs. semi-synthetic data



Huang et al., *Nature Methods* (2018)

Real vs. semi-synthetic data



Benchmark standard

		labels used in Huang *et al* .						
		0	**1**	**2**	**3**	**4**	**5**	**6**
labels reported in Zeisel *et al* .	**CA1-Pyramidal**	442	20	289	1	4	42	40
	S1-Pyramidal	2	273	1	1	0	32	11
	Oligodendrocytes	0	0	0	282	0	62	2
	Interneurons	5	7	2	0	220	6	1
	Endothelial	0	0	0	0	1	0	14
	Microglia	0	0	0	0	0	0	6
	Mural	0	1	0	0	0	0	0
	Ependymal	0	0	0	0	0	0	7
	Astrocytes	0	1	0	2	0	1	20

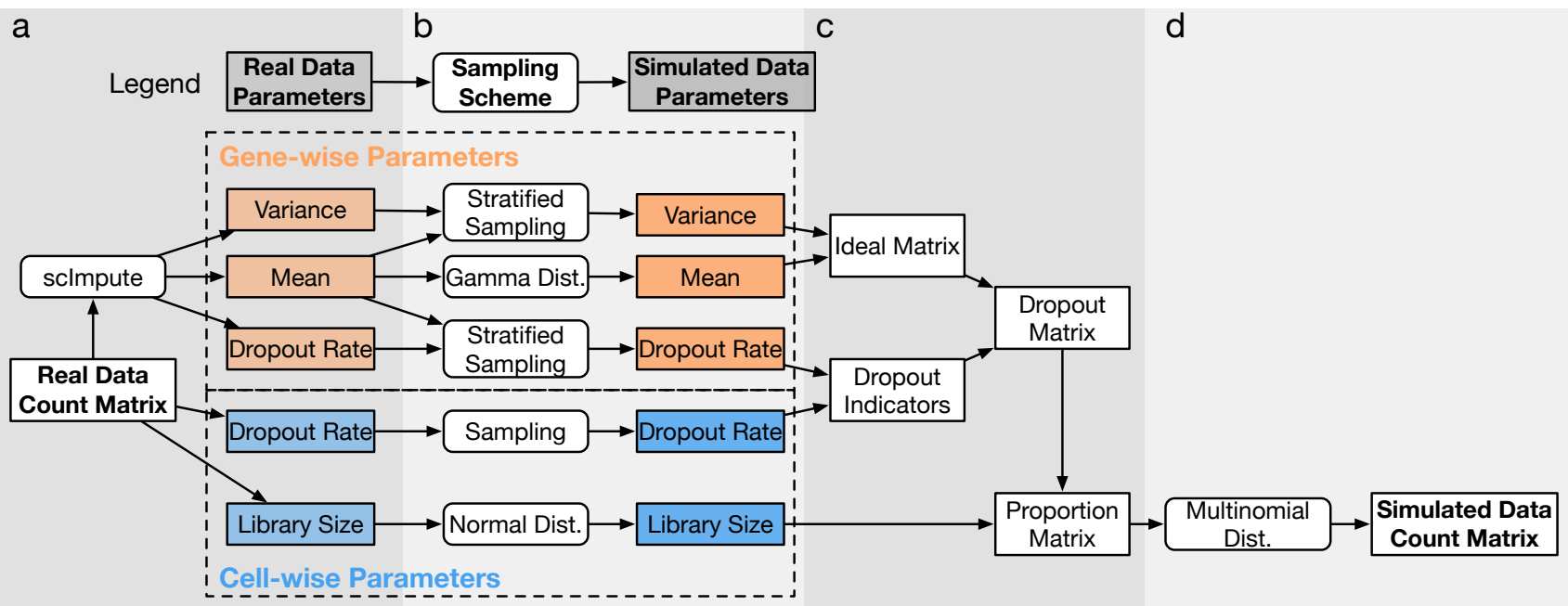
scDesign: statistical simulator for experimental design

Simulation-based scRNA-seq experimental design

Advantages of scDesign:

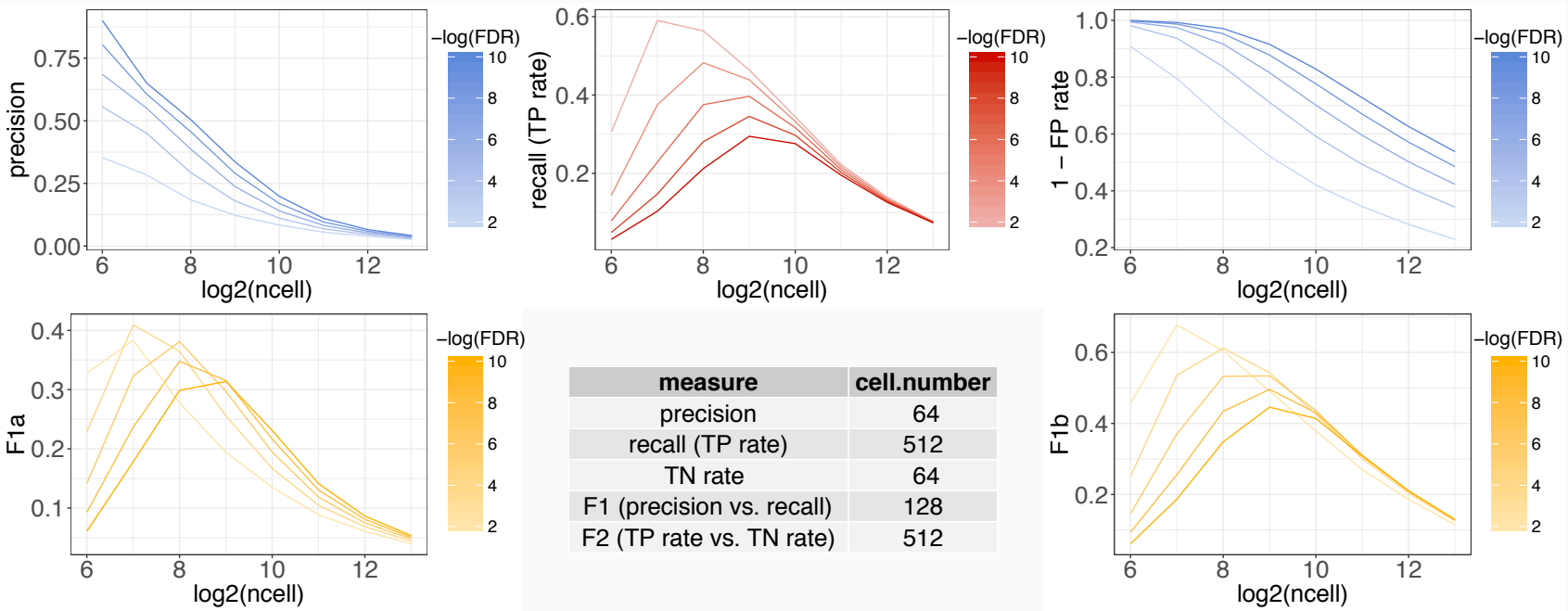
- Protocol-adaptive and data-adaptive: learn from
  - Public scRNA-seq data
  - Pilot-study data
- Generate synthetic data that well mimic real data under a pre-specified experimental setting
  - Assist experimental design & method development
- Flexible in accommodating user-specific analysis needs
- No experimental cost

Generative framework of scDesign



Experimental design for gene differential expression

Astrocytes vs. Oligodendrocytes (Fluidigm C1)



measure	cell.number
precision	64
recall (TP rate)	512
TN rate	64
F1 (precision vs. recall)	128
F2 (TP rate vs. TN rate)	512

scDesign paper

Bioinformatics, 35, 2019, i41–i50
doi: 10.1093/bioinformatics/btz321
ISMB/ECCB 2019

OXFORD

A statistical simulator scDesign for rational scRNA-seq experimental design

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*To whom correspondence should be addressed.

Summary

Bulk RNA-seq

- Isoform identification: **AIDE**
- Isoform quantification: **MSIQ**

Single-cell RNA-seq

- Dropout imputation: **scImpute**
- Simulator & experimental design: **scDesign**

Acknowledgements

Dr. Wei Vivian Li

former PhD student @UCLA

currently Assistant Professor @Rutgers

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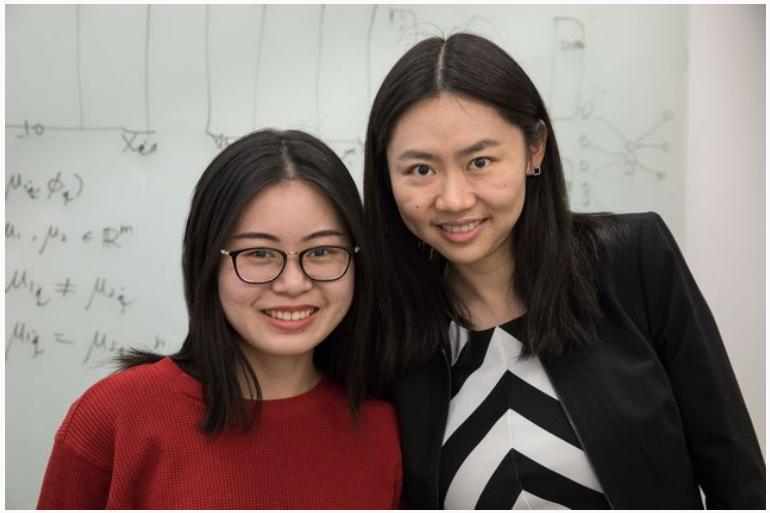
Dr. Xin Tong (USC)

Dr. Shihua Zhang (CAS)

Dr. Anqi Zhao (NUS)



Website: http://jsb.ucla.edu